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# Fish Welfare: the use of CO<sub>2</sub> to avoid fish suffering during preparation to processing

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**Key words:** antifreeze, CO<sub>2</sub>, carp, fish welfare, mullet, processing plant, striped bass

#### Abstract

Fish welfare has become a significant concern during the last decade. The issue of slaughter is getting the interest of many consumer organizations and fish growers, and suppliers. We need to take careful consideration to food fish slaughtering, and not only for an ethical reason. Numerous quality traits are in place already at the end of fish life. Nonetheless, other traits can be developed while the post mortem muscles are turning into consumer produce. Therefore, it is necessary to investigate ways to avoid the suffering of the fish before their processing. To improve the efficiency of the immobilizing and stunning process of the fish at the entrance of the fish plant, we investigated the use of  $CO_2$  in combination with low temperature. When combining  $CO_2$  to  $CO_2$  water temperature, fish were stunned and permanently immobilized within 20 min or less, while their core temperature was  $CO_2$  in the fish receiving tank of the processing plant might be more appropriate for animal welfare. Therefore, we suggest continuing our experiments in the fish processing plant.

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## Introduction

Nowadays, both fish consumers and aquaculture producers are concerned about animal welfare. Therefore, a request for an investigation was submitted to our laboratory via a Ministry of Agriculture and Rural Development Veterinarian Services representative, prompted by signs of life in a carp in the packing plant. The regulations require that the fish must be stunned before the sorting and processing starts, for welfare reason, while the fish's core temperature much not be higher than 4°C for concerns related to the consumer's health. It appears that the reported case was during midwinter, during which the average water temperature in the fish ponds is 12°C.

Despite the hot climate of the Northern Valleys, the fish center of the Israeli fish industry, the aquaculture farmers also produce fish species such as various carps and striped bass, which originate from the earth's middle latitudes (40° to 60° north and south of the Equator). Carp, for example, can live at the bottom of lakes in Europe submerged at 4°C during the winter. The routine procedures of permanently immobilizing fish before sorting and packing in the Israeli processing plants are hosting them in a 2°C pool of tap water for about 20 min. In contrast, the fish ponds' water temperature during the summer can reach 31°C. So naturally, fish species of northern origin, such as carp, die due to the over 20°C temperature immediate reduction.

The adaptation of fish to low temperatures is the result of long-term evolution. For example, amur carp (*Cyprinus carpio haematopterus*) survives low temperatures (0-4°C) for six months per year (Liang et al., 2015). Moreover, Liu and coworkers (2009) characterized the antifreeze protein during embryogenesis and hatched larvae in the gibel carp (*Carassius auratus gibelio*). Also, the precursor of the antifreeze protein concentration varies with the season; it is at its peak in November and lowest in May (Hew et al., 1999). Thus, it is not surprising that by introducing carps to a 2°C pool of tap water for about 20 min during the cold winter, they stay alive. Therefore, we had to consider adding a factor to the cold water that will help avoid the suffering of the fish before their sorting and packing process but will not affect the consumers' health or change the fish smell and taste.

In Israel, after harvesting the fishponds, the farmers transfer the fish to cool, oxygenated water tanks, carrying the fish to the 2°C water introductory tank in the process plant. As a result, the fish body temperature decreases rapidly, and the fish lose consciousness because of anoxia. This easy and quick procedure became the standard in Mediterranean Countries and the UK for rainbow trout (Poli et al., 2005). Since fish euthanized by chemicals should not be offered to humans,  $CO_2$  is considered a treatment that avoids unacceptable residues (American Veterinary Medical Association, 2020). Usually, the fish, such as salmon, turned unconscious within a few minutes (Robb, 2001; Roth et al., 2006; Wall, 2001). Wall (2001) offered to refine the method and modify it to speed up stunning using  $CO_2$ . Therefore, we investigated the use of  $CO_2$  to successfully avoid the suffering of the fish by shortening its immobilizing time.

### **Materials and Methods**

Fish

One hundred of each species, adult, ~500-gr common carp (*Cyprinus carpio*), mullet (Mugil cephalus) and hybrid striped bass (striped bass, *Morone saxatilis* crossed with white bass, *M. chrysops*) were supplied by the Nir David aquaculture farm. The fish were let to adapt in flow through 1000-L tanks, with water temperature of 18°C for a week.

## Experiment procedures

Fish of each species was divided into a set of 5-100L tanks, five fish per tank. Ten tanks had fish in 2°C water, five tanks with bubbling  $CO_2$  and five tanks without, and ten tanks with 20°C water temperature, 5 with bubbling  $CO_2$ , and five without bubbling  $CO_2$ . Carbon dioxide was bubbled at the bottom of the tanks via air stones at the flow rate of 20-L/min. After 20 min., the fish was placed on crushed ice, the fish's core temperature was sampled, and euthanasia was determined by macroscopic and behavior observations. The

observations during and after immersion included loss of equilibrium, stopping opercular movement, response to tail-fin pinch, and eye movement (Collymore et al., 2014).

## Carbon dioxide (CO<sub>2</sub>) measurements

Water samples were taken by submersing 500-ml clear sampling bottle that was capped close tight under the surface of the water. Temperature, pH, conductivity and alkalinity were measured with a few minutes and CO<sub>2</sub> values were calculated.

#### Ethic committee

The authors received certification to conduct the experiments mentioned here by the Ethic Committee of the Israeli Ministry of Agriculture and Rural Development. Each of the authors is a certified researcher by the Ethic Committee of the Agriculture Research Organization of the Israeli Ministry of Agriculture and Rural Development.

## Results

The results of our experiments are shown in **Table 1**. Three fish species, common carp, hybrid striped bass, and flathead grey mullet, were tested in at temperatures  $20^{\circ}\text{C}$  versus  $2^{\circ}\text{C}$ , with and without  $CO_2$ . Bubbling  $CO_2$  shortens the time of immobilizing and euthanizing while achieving full stillness of the fish. However, only a water temperature of  $2^{\circ}\text{C}$  in concert with  $CO_2$  achieved 100% success. This treatment also provided the needed fish's core temperature of less than  $4^{\circ}\text{C}$ .

<b>Table 1</b> Average values of CO <sub>2</sub> in the experimental tanks after 20 min.

	CO₂ Values (mg/L)			
	20°C air 1	20°C+CO <sub>2</sub> <sup>2</sup>	2°C+air <sup>3</sup>	2°C+CO2 4
Carp	8	1198	11	2149
Striped Bass	5	1553	8	1819
Mullet	4	1127	9	2043

<sup>&</sup>lt;sup>1</sup> No euthanized fish.

## **Discussion**

Poikilothermic (cold-blooded) animals have developed several biochemical and physiological adaptations to survive exposure to diurnal or seasonal low temperatures and chronic cold. Numerous investigations mentioned the adaptive changes for cold survival of the Antarctic notothenioid. A breakthrough identified antifreeze glycoprotein (AFGP) is considered one of the significant factors in expanding polar fish populations into harsh environments (Devries, 1971). Also, discovering the loss of many genes, including hemoproteins in the icefish family Channichthyidae, seems to respond to constant cold water (Sidell and O'Brien, 2006). However, eurythermal fish such as the common carp (Cyprinus carpio) use different strategies to survive low temperatures. One famous hypothesis regarding the survival mechanism at cold temperature is "homeoviscous adaptation," which suggests that fish can avoid freezing by decreasing the saturation of membrane phospholipids to offset the cold-induced rigidification of lipid bilayers during cold periods (Tiku et al., 1996).

Those fishes in low-temperature environments have evolved some unique physiological mechanisms to adapt to icy conditions (Chen et al., 2008; Fletcher et al., 2001; Kim et al., 2017). For example, the production of antifreeze protein, lack of hemoglobin, and tubulin synthesis can improve the adaptability of Antarctic fishes to low-temperature environments

<sup>&</sup>lt;sup>2</sup> 75% euthanized with ambient fish core temperature.

<sup>&</sup>lt;sup>3</sup> 67% euthanized with 3.5°C fish core temperature.

<sup>&</sup>lt;sup>4</sup> 100% euthanized with 3.5°C fish core temperature.

(Fletcher et al., 2001; Parker and Detrich, 1998). However, eurythermal freshwater fishes are exposed to a wide range of fluctuations in water temperature rather than in a relatively stable temperature environment. Therefore, eurythermic fish must adjust physiological processes quickly to respond to the acute cooling when severe cold fronts approach in winter. Interestingly enough, Raymond (1992) reported that glycerol is a colligative antifreeze in some northern fishes.

Carbon dioxide was first mentioned as a fish anesthetic since it could immobilize fish (Fish, 1943). Either bubbling  $CO_2$  gas can produce carbon dioxide in the fish tank or the soda-acid technique (Post, 1979; Summerfelt and Smith, 1990). It is safe for humans, and its use is unrestricted (Summerfelt and Smith, 1990).  $CO_2$  does not induce analgesia and has a "shallow effect" compared to MS-222 (Anderson et al., 1997). Physiological responses to  $CO_2$ , such as no decrease in blood PO2 and lowered plasma pH during anesthesia, differ from benzocaine, 2-phenoxyethanol, MS-222, and metomidate (Iwama et al., 1989). Acerete et al. (2009) show that both  $CO_2$  and asphyxia in ice are suitable for processing fish. However, the  $CO_2$  treatment showed lower lactate and cortisol response compared to asphyxia in ice. In terms of quality, differences between methods are minor. Cortisol concentration revealed that MS-222, clove oil, and  $CO_2$  did not cause significant stress to steelhead trout (Pirhonen and Schreck 2003). Our results suggest that combining low temperature with bubbling  $CO_2$  in the fish receiving tank of the processing plant might be a more satisfactory strategy for animal welfare.

Moreover, only a water temperature of  $2^{\circ}$ C in concert with  $CO_2$  achieved 100% successful results. It was the only treatment that provided the needed fish's core temperature of less than  $4^{\circ}$ C. Therefore, we suggest validating our results in a field study in the fish processing plant conditions.

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## References

Acerete, L., Reig, L., Alvarez, D., Flos, R. and Tort, L., 2009. Comparison of two stunning/slaughtering methods on stress response and quality indicators of European sea bass (*Dicentrarchus labrax*). *Aquaculture*, 287(1-2), pp.139-144. https://doi.org/10.1016/j.aquaculture.2008.10.012

**American Veterinary Medical Association,** AVMA Guidelines for the Euthanasia of Animals: 2020 Edition; American Veterinary Medical Association: Schaumberg, IL, USA, 2020. ISBN 978-1-882691-54-8

**Anderson, W.G., McKinley, R.S. and Colavecchia, M.,** 1997. The use of clove oil as an anesthetic for rainbow trout and its effects on swimming performance. North American Journal of Fisheries Management, 17(2), pp.301-307. https://doi.org/10.1577/1548-8675(1997)017<0301:TUOCOA>2.3.CO;2

Chen, Z., Cheng, C.H.C., Zhang, J., Cao, L., Chen, L., Zhou, L., Jin, Y., Ye, H., Deng, C., Dai, Z. and Xu, Q., 2008. Transcriptomic and genomic evolution under constant cold in Antarctic notothenioid fish. *Proceedings of the National Academy of Sciences*, 105(35), pp.12944-12949. https://doi.org/10.1073/pnas.0802432105

**Collymore, C., Tolwani, A., Lieggi, C. and Rasmussen, S.,** 2014. Efficacy and safety of 5 anesthetics in adult zebrafish (*Danio rerio*). *Journal of the American Association for Laboratory Animal Science*, *53*(2), pp.198-203.

**Devries, A.L.,** 1971. Glycoproteins as biological antifreeze agents in Antarctic fishes. *Science*, *172*(3988), pp.1152-1155. https://doi.org/10.1126/science.172.3988.1152

- **Fish, F.F.,** 1943. The anaesthesia of fish by high carbon-dioxide concentrations. Transactions of the American Fisheries Society, 72(1), pp.25-29.
- **Fletcher, G.L., Hew, C.L. and Davies, P.L.,** 2001. Antifreeze proteins of teleost fishes. *Annual review of physiology*, 63(1), pp.359-390.
- Hew, C., Poon, R., Xiong, F., Gauthier, S., Shears, M., King, M., Davies, P. and Fletcher, G., 1999. Liver-specific and seasonal expression of transgenic Atlantic salmon harboring the winter flounder antifreeze protein gene. *Transgenic Research*, 8(6), pp.405-414. https://doi.org/10.1023/A:1008900812864
- **Iwama, GK, McGeer, JC and Pawluk, MP,** 1989. The effects of five fish anaesthetics on acid base balance, hematocrit, blood gases, cortisol, and adrenaline in rainbow trout. Canadian Journal of Zoology, 67 (8), pp. 2065-2073. https://doi.org/10.1139/z89-294
- **Kim, H.J., Lee, J.H., Hur, Y.B., Lee, C.W., Park, S.H. and Koo, B.W.,** 2017. Marine antifreeze proteins: structure, function, and application to cryopreservation as a potential cryoprotectant. *Marine drugs*, *15*(2), p.27. https://doi.org/10.3390/md15020027
- **Liang, L., Chang, Y., He, X. and Tang, R.,** 2015. Transcriptome analysis to identify cold-responsive genes in amur carp (*Cyprinus carpio haematopterus*). *PLoS One*, *10*(6), p.e0130526. https://doi.org/10.1371/journal.pone.0130526
- **Liu, J.X., Zhai, Y.H. and Gui, J.F.,** 2009. Molecular characterization and expression pattern of AFPIV during embryogenesis in gibel carp (*Carassius auratus gibelio*). *Molecular biology reports*, 36(7), pp.2011-2018. https://doi.org/10.1007/s11033-008-9412-3
- **Parker, S.K. and Detrich III, H.W.,** 1998. Evolution, Organization, and Expression of α-Tubulin Genes in the Antarctic Fish Notothenia coriiceps: Adaptive Expansion of a Gene Family by Recent Gene Duplication, Inversion, and Divergence. *Journal of Biological Chemistry*, *273*(51), pp.34358-34369.
- **Pirhonen, J. and Schreck, C.B.,** 2003. Effects of anaesthesia with MS-222, clove oil and CO<sub>2</sub> on feed intake and plasma cortisol in steelhead trout (Oncorhynchus mykiss). *Aquaculture*, 220(1-4), pp.507-514. https://doi.org/10.1016/S0044-8486(02)00624-5
- **Poli, B.M., Parisi, G., Scappini, F. and Zampacavallo, G.,** 2005. Fish welfare and quality as affected by pre-slaughter and slaughter management. *Aquaculture International*, 13(1), pp.29-49.
- **Post, G.,** 1979. Carbonic acid anesthesia for aquatic organisms. *The Progressive Fish-Culturist*, *41*(3), pp.142-144. https://doi.org/10.1577/1548-8659(1979)41[142:CAAFAO]2.0.CO;2
- **Raymond, J.A.,** 1992. Glycerol is a colligative antifreeze in some northern fishes. *Journal of Experimental Zoology*, 262(3), pp.347-352. sci-hub.se/10.1002/jez.1402620316
- **Robb, D.H.F.,** 2001. The relationship between killing methods and quality. *Farmed fish quality*.
- **Roth, B., Slinde, E. and Robb, D.H.,** 2006. Field evaluation of live chilling with CO<sub>2</sub> on stunning Atlantic salmon (*Salmo salar*) and the subsequent effect on quality. *Aquaculture Research*, *37*(8), pp.799-804.

**Sidell, B.D. and O'Brien, K.M.,** 2006. When bad things happen to good fish: the loss of hemoglobin and myoglobin expression in Antarctic icefishes. *Journal of Experimental Biology*, 209(10), pp.1791-1802.

**Summerfelt, R.C.,** 1990. Anesthesia, surgery, and related techniques. Methods for fish biology. ISBN 0-913235-58-X

**Tiku, P.E., Gracey, A.Y., Macartney, A.I., Beynon, R.J. and Cossins, A.R.,** 1996. Cold-induced expression of  $\Delta 9$ -desaturase in carp by transcriptional and posttranslational mechanisms. *Science*, *271*(5250), pp.815-818. https://doi.org/ 10.1126/science.271.5250.815

**Wall, A.J.,** 2001. Ethical considerations in the handling and slaughter of farmed fish. Farmed fish quality. Blackwell Science. Bristol, England, pp.108-119.