





Original Research Articles

A First Look at Genetic Diversity of *Metapenaeus ensis* Populations in Tam Giang – Cau Hai Lagoon, Vietnam

Nguyen Xuan Huy^{1,2} , Nguyen Ty² , Tran Van Giang² , Tran Vinh Phuong¹ ¹ Department of Science, Technology and International relations, Hue University, ² Faculty of Biology, University of Education, Hue UniversityKeywords: *Metapenaeus ensis*, mtCOI, Tam Giang – Cau Hai lagoon, shrimp, biodiversity, population structure<https://doi.org/10.46989/001c.117579>

Israeli Journal of Aquaculture - Bamidgheh

Vol. 76, Issue 2, 2024

This study investigates the genetic diversity and population structure of the greasyback shrimp, *Metapenaeus ensis* (De Haan, 1844), within the Tam Giang – Cau Hai lagoon, Vietnam, by analyzing mtCOI genes from 91 individuals collected across four populations in nine sampling locations. High genetic diversity was found, with 34 unique haplotypes and 38 genetic variations identified. Most genetic variation occurred within populations (AMOVA), suggesting high gene flow. Low and non-significant *F*_{st} values and close genetic distances confirmed minimal differentiation among populations. These findings provide the first insights into *M. ensis* population genetics in this lagoon, informing conservation and management efforts. Understanding genetic diversity is crucial for sustainable management and conservation of marine species. This research aids in understanding population resilience, potential impacts of overfishing, and may assist in developing sustainable harvest strategies of this economically important shrimp species.

INTRODUCTION

Metapenaeus ensis, commonly known as the greasyback shrimp or sand shrimp, is one member of the family Penaeidae.¹ The shrimp is a commercially valuable species native to the Indo-West Pacific region. Widely distributed throughout the Indian and West Pacific Oceans, it's found in areas like Sri Lanka, Malaysia, southeast China, Japan, and northern Australia.^{1,2} Beyond its wild populations, *M. ensis* is also a prominent species in aquaculture, particularly across Southeast Asia. Countries like Malaysia, Indonesia, Thailand, Philippines, Taiwan, and Vietnam all have established traditions of cultivating this shrimp.³ In Vietnam, *M. ensis* thrives in various coastal regions, including Hai Phong, Quang Ngai, Da Nang, Thua Thien Hue, Quang Tri, and Quang Binh.^{4,5} One particularly important habitat for *M. ensis* in Vietnam is the Tam Giang – Cau Hai lagoon system. Located within Thua Thien Hue province, this lagoon complex holds the distinction of being the largest semi-closed lagoon system in all of Southeast Asia. Stretching over a vast 68 kilometers and covering an impressive 22,000 hectares, the lagoon is further divided into four distinct sub-lagoons progressing north to south: Tam Giang lagoon, Sam lagoon, Ha Trung - Thuy Tu lagoon, and Cau Hai lagoon. The Tam Giang – Cau Hai lagoon system boasts exceptional biodiversity, flourishing with a diverse array of ecosystems. Over 1,296 species have been identified within the lagoon, with a remarkable 41 classified as

rare and precious.^{6,7} *M. ensis* plays a vital role within the Tam Giang – Cau Hai ecosystem. Local communities heavily rely on harvesting this shrimp to supply the food chain and generate income, thanks in part to the high quality of its meat. Various fishing techniques, including barrier nets, cast nets, trap nets, and bag nets, are employed to capture *M. ensis*.

DNA barcoding, a technology utilizing short, standardized DNA sequences, is a powerful tool for assessing biodiversity, identifying species, and understanding evolutionary relationships (phylogeny).⁸ It relies on analyzing a specific gene segment, the *cytochrome oxidase subunit I* (COI), to distinguish between species. This technique has proven valuable in various studies, including evaluating biodiversity in a mangrove hotspot,⁹ assessing the diversity of freshwater shrimp in Henan, China¹⁰ and identifying commercially important marine shrimp in India.¹¹ Furthermore, the complete mitochondrial genome of *M. ensis* has been sequenced and deposited in a public database.¹² DNA barcoding plays a critical role in furthering our understanding of shrimp biodiversity. It provides a reliable and efficient method for species identification, population differentiation and informing conservation efforts.

The Tam Giang – Cau Hai lagoon stands as the primary hub for shrimp production and fishing in central Vietnam. The adverse impacts of climate change are keenly felt, particularly in coastal regions, affecting human lives and livelihoods.^{13,14} Various manifestations of climate change, in-

a Corresponding author. Tel.: +84 912734565, e-mail: nguyenxuanhuy@hueuni.edu.vn

Table 1. Sampling location of individual *M. ensis* in Tam Giang – Cau Hai Lagoon

No.	Coordinate	Sampling location	Population	Sample
1	16°38'12"N 107°28'37"E	Northern Tam Giang lagoon (Dien Hai - Quang Ngan - Quang Cong)	Tam Giang lagoon (TG)	10
2	16°34'9"N 107°35'39"E	Southern Tam Giang lagoon (Hai Duong - Huong Phong - Thuan An)		11
3	16°30'8"N 107°39'15"E	Sam lagoon (Phu An – Phu My)	Sam – Ha Trung – Thuy Tu lagoon (ST)	10
4	16°28'39"N 107°43'38"E	Ha Trung lagoon (Phu Xuan – Phu Da)		10
5	16°24'6"N 107°48'15"E	Thuy Tu lagoon (Vinh An – Vinh Hung)		10
6	16°20'23"N 107°53'6"E	Eastern Cau Hai lagoon (Phu Loc)	Cau Hai lagoon (CH)	10
7	16°19'4"N 107°48'26"E	Western Cau Hai lagoon (Vinh Giang – Vinh Hien)		10
8	16°15'20"N 108° 2'38"E	Northern Lang Co lagoon (An Cu Dong)	Lang Co lagoon (LC)	10
9	16°13'22"N 108° 3'14"E	Southern Lang Co lagoon (An Cu Tay)		10

cluding escalating temperatures, sea-level rise, diseases, harmful algal blooms, alterations in rainfall patterns, uncertainties in external input supplies, changes in sea surface salinity, and severe climatic events, can profoundly influence the natural resources of *M. ensis* in aquatic ecosystems. This study contributes to this growing body of knowledge by providing the first data on the genetic diversity and population structure of *M. ensis* within the Tam Giang – Cau Hai lagoon. We sequenced and analyzed the COI gene region of 91 individuals collected from four distinct locations within the study area, expanding the existing COI sequence database for this species.

MATERIALS AND METHODS

SAMPLE COLLECTION

Specimens of *M. ensis* were collected from the Tam Giang – Cau Hai lagoon between February 2022 and December 2023 (Table 1). Two methods were employed: direct capture and acquisition from local fishermen within the sampling region. To maintain specimen integrity for morphological analysis (shaping and photographing), shrimp were immediately placed in an icebox and transported swiftly to the laboratory. For subsequent DNA extraction, one gram of muscle segment or a single whole leg (pereopod) was excised from each specimen and stored at -80°C.

GENOMIC DNA EXTRACTION, PCR AND SEQUENCING

One gram of muscle segment or a single whole leg (pereopod) was ground in liquid nitrogen using a mortar and pestle. Total DNA was extracted following the manufacturer's protocol for the GeneJET Genomic DNA Purification Kit (Thermo Fisher Scientific). Genomic DNA concentrations were quantified using a Nanodrop 2000 spectrophotometer

(Thermo Fisher Scientific Inc., USA) and stored at 4 °C until further analysis.

The *mtCOI* gene region was amplified using polymerase chain reaction (PCR) with specifically designed primers based on the *M. ensis* complete mitochondrial genome (Accession numbers: MK500697.1, NC_026834.1 and ON599337.1) and a previous study.⁹ The primers were: forward 5'-CTTGCAGGGGTCTCATCAAT-3' and reverse 5'-GC-GAAGATCCCCAAATACAGC-3'. PCR reaction contained 0.2 µg genomic DNA, 10 pmol primers, 12.5 µL (2X) PCR Master Mix (Thermo Fisher Scientific) in a total volume of 25 µL. The PCR reaction was conducted with the following PCR condition: 1 cycle at 95 °C for 5 min; 30 cycles at 95 °C for 30 s, 55 °C for 30s, and 72 °C for 30s; and 1 cycle at 72 °C for 5 min. The resulting PCR amplicons were visualized on a 1.0% agarose gel stained with Safe-dye™ and examined under UV light. Sanger DNA sequencing was subsequently performed at First Base (Malaysia) using the Capillary Electrophoresis method by using the above PCR primers.

SEQUENCING ANALYSIS AND NCBI DATABASE'S REGISTRATION

The obtained COI sequences were extracted from chromatogram files after low quality bases were trimmed, and the forward and reverse sequences were joined into contigs aligned using the MUSCLE algorithm in the MEGA 11 Suite¹⁵ and edited using BioEdit 7.2.5 software.¹⁶ The nucleotide sequences were used as queries to search for homologous sequences in the NCBI database using BLASTn. The homologous sequences were used as references for comparison. Finally, the sequences were registered in the NCBI database with accession PP348174 to PP348207.

Table 2. Similarity level and query coverage of *mtCOI* gene segments when using Blast search on NCBI with reference of MK500697.1, NC_026834.1, ON599337.1.

Haplotype (Hap)	Similarity (%)	Query Coverage (%)
21	98.85	100
27	98.99	100
9; 13	99.14	100
2; 10; 15; 18; 22; 33	99.28	100
4; 20; 29; 32	99.42	100
3; 5; 7; 14; 26; 30	99.57	100
1; 6; 8; 12; 16; 17; 19; 23; 24; 25; 28; 31; 34	99.71	100
11	99.86	100

GENETIC DIVERSITY ANALYSIS

To analyze the genetic diversity, various indices were calculated using DnaSP v6.10.01 software¹⁷: number of variable sites (S), number of mutations (η), average number of nucleotide differences (k), number of haplotypes (h), nucleotide diversity (π), haplotype diversity (Hd), Tajima's D, and Fu's Fs. We used Fu's Fs¹⁸ and Tajima's D¹⁹ to assess population expansion patterns in *M. ensis*. Fu's Fs analyzes the distribution of haplotypes, while Tajima's D compares pairwise differences among sequences to estimate allele frequency.²⁰ We determined the genetic differentiation index (Fst) and evaluated genetic differences within and among populations using analysis of molecular variance (AMOVA) with Arlequin software.²¹ Lastly, a Bayesian phylogenetic tree was reconstructed with BEAST version 2.7,²² employing the HKY substitution model and the Coalescent Bayesian Skyline tree prior. The Markov chain Monte Carlo (MCMC) search was set at 1,000,000 generations with burn-in at 10%. The iTOL web tool (<https://itol.embl.de/>) was used to visualize and refine the phylogenetic trees.

RESULTS

GENETIC POLYMORPHIC SPECIALTY OF *M. ENSIS* IN THUA THIEN HUE

Specimens of greasyback shrimp (*M. ensis*) were collected from 09 different places of Tam Giang – Cau Hai lagoon and were divided into 04 populations (Table 1). A total of 91 samples were preserved and extracted total genomic DNA. The COI amplicons were visualized by Agarose electrophoresis and sequenced with the size of 695 bp. The obtained sequences were aligned with sequences on GenBank using the BLAST search shown that *M. ensis* samples collected in Tam Giang – Cau Hai lagoon had a very-high similarity rate of 98.85 % - 99.86 % in compared to with 3 references COI fragment sequence sequences MK500697.1, NC_026834.1, ON599337.1 (Table 2).

GENETIC DIVERSITY

Genetic diversity is the biological variation occurring within a species to make it possible for species to adapt

with the environment changes. Analysis of 91 individuals gathered from the studied area, 34 unique haplotypes and 38 polymorphic sites were defined. Thirty four haplotypes were registered on GenBank with the accession number from PP348174 to PP348207. A total of 62 variable sites (S) and 39 mutations were identified. The nucleotide diversity (π) was determined to be 0.00517 ± 0.00036 , while the haplotype diversity (Hd) was 0.884 ± 0.023 (Table 3). Notably, across all sampling locations, the Hd value varied, with the lowest and highest values observed in samples from LC and TG, respectively. Specifically, samples obtained from TG showed the highest Hd (0.938 ± 0.039), while the lowest Hd (0.842 ± 0.061) was observed in samples collected from LC. In terms of π , LC exhibited the highest value (0.00588 ± 0.00070), whereas the lowest value (0.00489 ± 0.00064) was recorded in samples obtained from ST (Table 3). The results showed that the greasyback shrimp (*M. ensis*) exhibited high haplotype diversity and nucleotide diversity.

GENETIC STRUCTURE

The AMOVA analysis revealed that genetic variation within *M. ensis* populations was the primary source of diversity, accounting for 100.73%. In contrast, genetic differentiation among groups (i.e., sampling sites) was negligible and statistically insignificant at -0.73% (Table 4). Furthermore, the low and statistically insignificant fixation index (Fst = -0.0073, P = 0.712) corroborated the lack of substantial genetic differences between the *M. ensis* populations sampled from different lagoons across Thua Thien Hue, Vietnam. Pairwise Fst comparisons further support this (range: -0.02177 to -0.00439, Table 5). In most populations, Fu's Fs and Tajima's D values did not indicate significant population expansion, with the exception of TG and ST sites for Fu's Fs (Table 3). Overall, these findings suggest minimal genetic differentiation among the *M. ensis* populations within the Tam Giang-Cau Hai lagoon system.

GENETIC DISTANCE

Genetic distance is a measure of the genetic divergence between species or between populations within a species, whether the distance measures time from common ancestor or degree of differentiation.²³ Evaluations were made on

Table 3. Genetic diversity of *M. ensis* populations in Tam Giang – Cau Hai Lagoons based - on *mtCOI* sequence

Population	TG	ST	CH	LC	Total
Number of samples	21	30	20	20	91
Number of variable sites (S)	19	17	18	19	38
Number of mutation (n)	19	17	18	19	39
Average number of nucleotide differences (k)	3.5429	3.4000	3.7474	4.0895	3.69492
Nucleotide diversity (per site) ($\pi \pm$ SD)	0.00510 \pm 0.00066	0.00489 \pm 0.00064	0.00539 \pm 0.00089	0.00588 \pm 0.00070	0.00517 \pm 0.00036
Number of Haplotypes (h)	14	14	11	10	34
Haplotype (gene) diversity (Hd \pm SD)	0.938 \pm 0.039	0.879 \pm 0.043	0.900 \pm 0.044	0.842 \pm 0.061	0.884 \pm 0.023
Tajima's D	-1.23047 (0.121)	-0.71052 (0.251)	-0.98346 (0.172)	-0.89403 (0.172)	-0.95462 (0.179)
Fu's Fs statistic	-6.818* (0.001)	-4.604* (0.020)	-2.873 (0.074)	-1.562 (0.199)	-3.96439 (0.0735)

*Significant statistic ($P < 0.05$)**Table 4. Analysis of Molecular Variance (AMOVA) results for *M. ensis* populations collected in Tam Giang – Cau Hai lagoon**

Source	Degree of freedom	Sum of squares	Variance components	Percentage of total variance (%)
Among populations	3	1.116	-0.00322 Va	-0.73
Within populations	87	38.664	0.44442 Vb	100.73
Total	90	39.780	0.44120	
Fixation index (Fst)	-0.00730	P = 0.71163 \pm	0.01577	

Table 5. Fst values (below dash) and probability values (above dash) among different *M. ensis* populations in Tam Giang – Cau Hai Lagoon

Population	TG	ST	CH	LC
TG	-	0.67871	0.77734	0.21289
ST	-0.01039	-	0.96875	0.49707
CH	-0.01583	-0.02177	-	0.34082
LC	0.01086	-0.00439	0.00451	-

91 sequences to determine the evolutionary divergence between nucleotide sequences of COI gene segments. The findings indicated that the *M. ensis* samples collected in Tam Giang – Cau Hai lagoon exhibited significant genetic similarities (Table 3). In addition, the *M. ensis* population in the studied area demonstrated a high level of genetic diversity among individuals, with genetic distances ranging from

0.00490 to 0.00549. In general, there were no substantial disparities in genetic distance between populations when comparing pairs. Genetic distance was close genetic inter-connectedness among the populations.

Based on the sequence of COI segments of collected individuals, the phylogenetic tree was built using Bayesian phylogenetic tree reconstruction method by Beast 2.7 (Figure

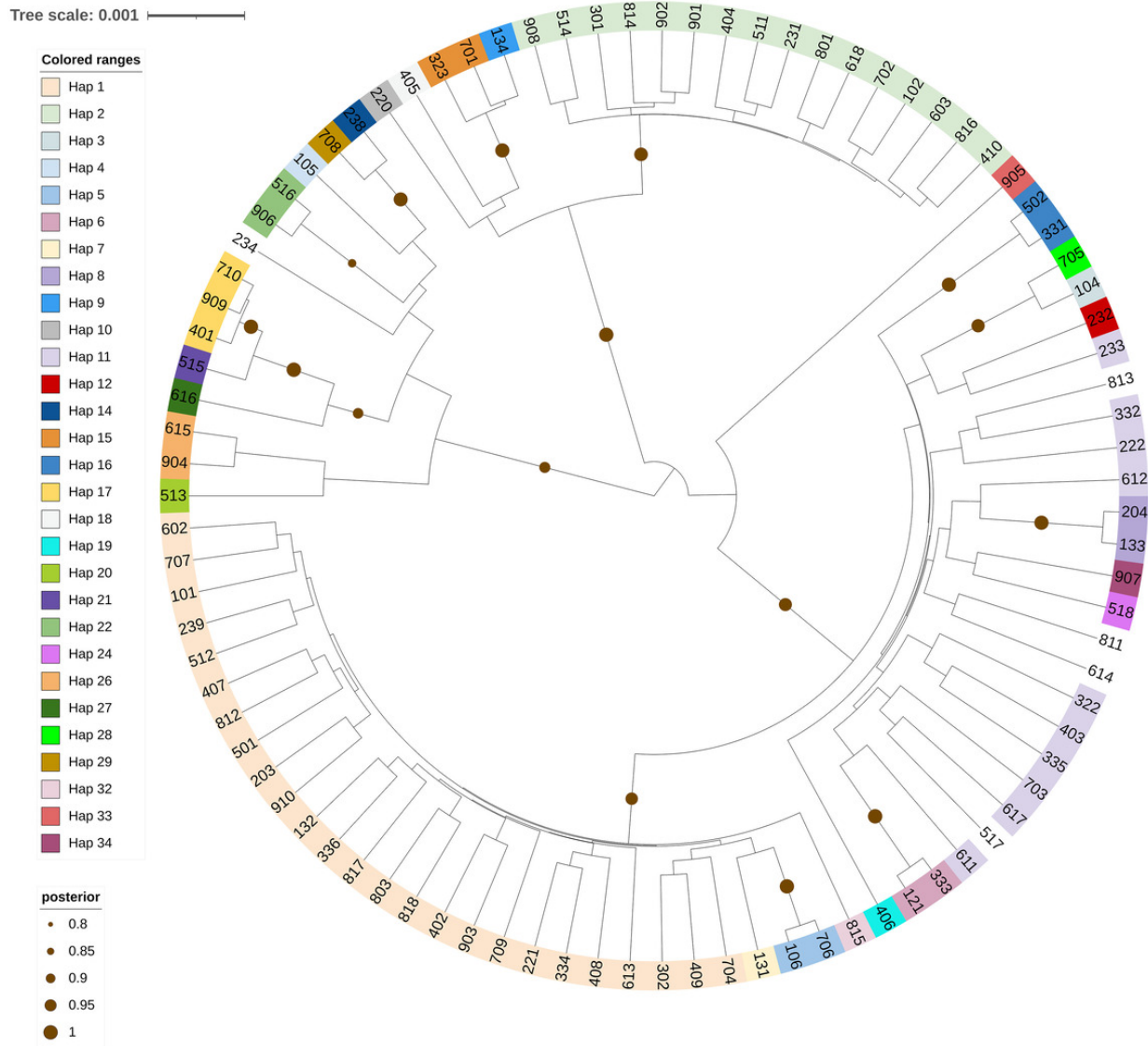


Figure 1. Phylogenetic tree of *M. ensis* population in Tam Giang - Cau Hai lagoon based on *mtCOI* amplicons using Bayesian phylogenetic tree reconstruction method by Beast 2.7.

1). The phylogenetic tree showed a total of 34 haplotypes were detected in 91 *M. ensis* sequences. Haplotype Hap 1, with observed frequently, was classified as the largest clade of the tree and was prevalent in samples collected from 4 distinct lagoons, namely TG, ST, CH and LC. Haplotype Hap 2 emerged as the second most predominant haplotype found in samples from these lagoons and grouped into the second largest clade. Haplotype Hap 11 also exhibited a high frequency in samples from TG, ST, and CH, located in the first largest clade with Hap 1, along with several other individual haplotypes. The post-test probability supporting the clades were quite high, demonstrating the close kinship between the studied populations.

DISCUSSION

DNA barcoding is a potent technique employed in species identification and exploration. It relies on standardized short DNA segments such as the *cytochrome oxidase subunit*

I (COI) gene for animals, and for plants: *rbcL*, *matK*, and *ITS*, as well as *ITS* for fungi, and the *16S rRNA* gene for bacteria and archaea. This method establishes a reference DNA barcode library.^{8,24} This technique has been widely applied in various domains, including biodiversity research, conservation, management of aquatic ecosystems, and ensuring the accuracy of seafood labeling and quality.^{25,26} Notably, the mitochondrial COI gene can form the cornerstone of a global bioidentification system for animals.²⁶ Comparative analyses utilizing GenBank BLASTn and BOLD search engines consistently demonstrate the COI gene's efficacy in identifying species with authenticated reference sequence data.²⁷ The fragment of COI gene has been efficiently used for shrimp species identification and genetic diversity.^{9-11,28,29} The determination of sample size and selection of sampling locations are critical tasks.³⁰ Prasertlux et al. conducted a study on COI polymorphism, analyzing 77 samples.³¹ Additionally, Shokoohmand et al. collected and analyzed 60 samples of wild white shrimp (*M. affinis*)

for diversity.²⁸ Similarly, Jamaluddin et al. utilized 74 shrimp specimens of the COI gene to assess patterns of DNA barcode variation in mangrove biodiversity.⁹ In our study, 91 samples from 04 different populations were collected and analyzed for genetic diversity, matching the sample size of the aforementioned studies.

The Tam Giang – Cau Hai lagoon system encompasses diverse aquatic vegetation, river estuaries, and mangrove forests, fostering a habitat for both aquatic and terrestrial life forms. The lagoon also supports various species of high economic value, such as shrimp and fish, which contribute to the socio-economic development of the region. Aquaculture in the lagoon plays a significant role in the socio-economic development of the region, providing livelihoods for many local residents.^{6,7,32,33} In a previous study, Nhung et al. (2015) distinguished between three types of aquaculture practices in the region: Earth ponds, net/bamboo stake ponds, and net enclosures.³⁴ The increasing amount of aquaculture structures reduced the water circulation, causing a build-up of contaminants in the lagoon.³⁴ Since 1995, there has been a notable deterioration in the environmental condition, leading to proposals for safeguarding the lagoon through the construction of wastewater treatment facilities.³⁵ The adverse impacts of climate change on shrimp production and aquaculture are evident.^{13,14,34} Genetic diversity analysis based on the *mtCOI* gene segment could provide crucial data for proposing measures to conserve aquatic resources.

M. ensis is celebrated for its remarkable productivity and delicious meat quality, recognized as a delicacy in Thua Thien Hue province, Vietnam. Research in the Tam Giang – Cau Hai lagoon has centered on various penaeid prawn species⁴ and the histological structure of the male gonad in greasyback shrimp.³⁶ In assessing the genetic diversity and population structure of *M. ensis*, analysis of partial COI sequences from populations in the Tam Giang Cau Hai lagoon revealed 34 haplotypes and 38 polymorphic sites (Table 1). The relatively robust genetic diversity observed in the *M. ensis* population of Tam Giang – Cau Hai lagoon (Tables 2 and 3) may be related to the random introduction of individuals from diverse spawning grounds or adaptations to local environmental factors and living conditions in Thua Thien Hue. Some individuals exhibited a tendency towards divergence compared to the overall evolutionary trajectory of the *M. ensis* population in Tam Giang – Cau Hai lagoon, possibly due to variations in habitat and geographical context. Genetic differentiation among populations is considered weak when the *Fst* value ranges from 0 to 0.05 (Table 5). Similarly, relative weakness in genetic differentiation occurs when *Fst* falls between 0.05 and 0.15. However, when *Fst* surpasses 0.15, genetic differentiation reaches a notably high level.³⁷ The substantial genetic diversity observed in the studied *M. ensis* populations suggests robust genetic interchange among shrimp populations within the lagoon system, facilitated by flats and the drift of larvae from spawning grounds into lagoons via estuaries, as well as the dynamic flow patterns within the lagoon.

CONCLUSIONS

The Tam Giang – Cau Hai lagoon, a recognized biodiversity hotspot in Southeast Asia, sustains diverse species and local food sources. Greasyback (*M. ensis*) shrimp, a commercially important delicacy, prompted a survey to assess its diversity within the lagoon due to concerns over potential overexploitation. This study presents the first data on greasyback shrimp diversity, revealing high levels of genetic variation with 34 haplotypes identified among 94 individuals. These findings provide insights into population structure and offer a crucial foundation for future research and potential conservation efforts aimed at ensuring the sustainability of this resource and the lagoon ecosystem.

ACKNOWLEDGMENTS

This work was supported by grant from the Ministry of Education and Training of Vietnam, under the grant number B2022-DHH-14 and Hue University, under the Core Research Program, Grant No. NCM.DHH.2022.07. [Tran Vinh Phuong] was funded by the Master, PhD Scholarship Programme of Vingroup Innovation Foundation (VINIF), code [VINIF.2023.TS.089].

AUTHORS' CONTRIBUTION

Conceptualization: Nguyen Xuan Huy (Equal), Nguyen Ty (Equal), Tran Van Giang (Equal), Tran Vinh Phuong (Equal). Data curation: Nguyen Xuan Huy (Equal), Nguyen Ty (Equal), Tran Van Giang (Equal), Tran Vinh Phuong (Equal). Formal Analysis: Nguyen Xuan Huy (Equal). Funding acquisition: Nguyen Xuan Huy (Lead). Investigation: Nguyen Xuan Huy (Equal), Nguyen Ty (Equal), Tran Van Giang (Equal), Tran Vinh Phuong (Equal). Methodology: Nguyen Xuan Huy (Equal), Nguyen Ty (Equal). Project administration: Nguyen Xuan Huy (Lead). Resources: Nguyen Xuan Huy (Equal), Nguyen Ty (Equal). Software: Nguyen Xuan Huy (Equal), Nguyen Ty (Equal). Supervision: Nguyen Xuan Huy (Lead). Validation: Nguyen Xuan Huy (Lead). Visualization: Nguyen Xuan Huy (Lead). Writing – original draft: Nguyen Xuan Huy (Lead), Nguyen Ty (Supporting). Writing – review & editing: Nguyen Xuan Huy (Lead).

COMPETING OF INTEREST – COPE

No competing interests were disclosed

ETHICAL CONDUCT APPROVAL – IACUC

The shrimp collection and experimental protocols followed the guideline from the Animal ethics committee of Hue University (<https://huaf.edu.vn/animalethics/>).

INFORMED CONSENT STATEMENT

All authors and institutions have confirmed this manuscript for publication.

DATA AVAILABILITY STATEMENT

All are available upon reasonable request.

Submitted: April 30, 2024 CST. Accepted: May 13, 2024 CST.

Published: May 16, 2024 CST.



This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CCBY-4.0). View this license's legal deed at <http://creativecommons.org/licenses/by/4.0> and legal code at <http://creativecommons.org/licenses/by/4.0/legalcode> for more information.

REFERENCES

1. Holthuis LB. FAO species catalogue. Vol. 1. Shrimps and prawns of the world. An annotated catalogue of species of interest to fisheries. *FAO Fish Synop.* 1980;125:1-126.
2. Chu KH, Chen QC, Huang LM, Wong CK. Morphometric analysis of commercially important penaeid shrimps from the Zhujiang estuary, China. *Fisheries Research.* 1995;23:83-93.
3. Viet TV, Sakuramoto K. Population structure and fishing of the greasyback shrimp (*Metapenaeus ensis*, De Haan, 1844) by bag net in a coastal river of the Mekong Delta, Vietnam. *International Journal of Fisheries and Aquaculture.* 2013;5:83-91.
4. Chat TT, To PTH, Mao ND, Chung NV. Research on species of penaeid prawns in coastal central provinces of Vietnam. *Journal of Fisheries Science and Technology.* 2009;1:31-36.
5. Thuan NV, Ha CTT. The species composition of prawn (Penaeidae) in the coastal zone of Quang Ngai Province. *Hue University Journal of Science.* 2008;49:177-181.
6. Van Tuyen T, Armitage D, Marschke M. Livelihoods and co-management in the Tam Giang lagoon, Vietnam. *Ocean & Coastal Management.* 2010;53(7):327-335. doi:[10.1016/j.ocecoaman.2010.04.001](https://doi.org/10.1016/j.ocecoaman.2010.04.001)
7. Virdis SG. An object-based image analysis approach for aquaculture ponds precise mapping and monitoring: a case study of Tam Giang-Cau Hai Lagoon, Vietnam. *Environ Monit Assess.* 2014;186(1):117-133. doi:[10.1007/s10661-013-3360-7](https://doi.org/10.1007/s10661-013-3360-7)
8. Kress WJ, Erickson DL. DNA barcodes: genes, genomics, and bioinformatics. *Proc Natl Acad Sci U S A.* 2008;105(8):2761-2762. doi:[10.1073/pnas.0800476105](https://doi.org/10.1073/pnas.0800476105)
9. Jamaluddin JAF, Mohammed Akib NA, Ahmad SZ, Abdul Halim SAA, Abdul Hamid NK, Mohd Nor SA. DNA barcoding of shrimps from a mangrove biodiversity hotspot. *Mitochondrial DNA A DNA Mapp Seq Anal.* 2019;30(4):618-625. doi:[10.1080/24701394.2019.1597073](https://doi.org/10.1080/24701394.2019.1597073)
10. Zhou CJ, Feng MX, Tang YT, Yang CX, Meng XL, Nie GX. Species diversity of freshwater shrimp in Henan Province, China, based on morphological characters and COI mitochondrial gene. *Ecol Evol.* 2021;11(15):10502-10514. doi:[10.1002/ece3.7855](https://doi.org/10.1002/ece3.7855)
11. Karuppasamy PK, Logeshwaran V, Priyadarshini RSS, Ramamoorthy N. Molecular and Systematic Identification of Food Marine Shrimps Using mtCOI Marker from Southeast Coast of India. *Thalassas: An International Journal of Marine Sciences.* 2020;36(2):487-495. doi:[10.1007/s41208-020-00201-3](https://doi.org/10.1007/s41208-020-00201-3)
12. Li Y, Falin Zhou, Oibin Yang, et al. Complete Mitochondrial Genome of *Metapenaeus Affinis* (H. Milne Edwards, 1837) and *Metapenaeus Ensis* (De Haan, 1844). *Israeli Journal of Aquaculture - Bamidgeh.* 2022;74:1-8. doi:[10.46989/001c.39657](https://doi.org/10.46989/001c.39657)
13. Maulu S, Hasimuna OJ, Haambiya LH, et al. Climate Change Effects on Aquaculture Production: Sustainability Implications, Mitigation, and Adaptations. *Frontiers in Sustainable Food Systems.* 2021;5. doi:[10.3389/fsufs.2021.609097](https://doi.org/10.3389/fsufs.2021.609097)
14. Yuniartik M, Setyaningrum EW, Yuniari SH, Faturakhmat SR, Prasetyo H. Climate change impact on shrimp (*Litopenaeus vannamei*) farming in Banyuwangi, East Java. *IOP Conference Series: Earth and Environmental Science.* 2022;1036(1):012062. doi:[10.1088/1755-1315/1036/1/012062](https://doi.org/10.1088/1755-1315/1036/1/012062)
15. Tamura K, Stecher G, Kumar S. MEGA11: Molecular Evolutionary Genetics Analysis Version 11. *Molecular Biology and Evolution.* 2021;38(7):3022-3027. doi:[10.1093/molbev/msab120](https://doi.org/10.1093/molbev/msab120)
16. Hall TA. *BioEdit: A User-Friendly Biological Sequence Alignment Editor and Analysis Program for Windows 95/98/NT.* Vol 41. Nucleic acids symposium series; 1999:95-98.
17. Rozas J, Ferrer-Mata A, Sánchez-DelBarrio JC, et al. DnaSP 6: DNA Sequence Polymorphism Analysis of Large Data Sets. *Mol Biol Evol.* 2017;34(12):3299-3302. doi:[10.1093/molbev/msx248](https://doi.org/10.1093/molbev/msx248)
18. Fu YX. Statistical Tests of Neutrality of Mutations Against Population Growth, Hitchhiking and Background Selection. *Genetics.* 1997;147(2):915-925. doi:[10.1093/genetics/147.2.915](https://doi.org/10.1093/genetics/147.2.915)
19. Tajima F. Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics.* 1989;123(3):585-595. doi:[10.1093/genetics/123.3.585](https://doi.org/10.1093/genetics/123.3.585)
20. Ramírez-Soriano A, Ramos-Onsins SE, Rozas J, Calafell F, Navarro A. Statistical Power Analysis of Neutrality Tests Under Demographic Expansions, Contractions and Bottlenecks With Recombination. *Genetics.* 2008;179(1):555-567. doi:[10.1534/genetics.107.083006](https://doi.org/10.1534/genetics.107.083006)

21. Excoffier L., Lischer HEL. Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources*. 2010;10(3):564-567. doi:[10.1111/j.1755-0998.2010.02847.x](https://doi.org/10.1111/j.1755-0998.2010.02847.x)
22. Bouckaert R, Heled J, Kühnert D, et al. BEAST 2: A Software Platform for Bayesian Evolutionary Analysis. *PLOS Computational Biology*. 2014;10(4):e1003537. doi:[10.1371/journal.pcbi.1003537](https://doi.org/10.1371/journal.pcbi.1003537)
23. Nei M, Maloy S, Hughes K. Genetic Distance. In: Maloy S, Hughes K, eds. *Brenner's Encyclopedia of Genetics (Second Edition)*. Academic Press; 2013:248-250. doi:[10.1016/B978-0-12-374984-0.00615-X](https://doi.org/10.1016/B978-0-12-374984-0.00615-X)
24. Antil S, Abraham JS, Sripoorna S, et al. DNA barcoding, an effective tool for species identification: a review. *Molecular biology reports*. 2023;50(1):761-775. doi:[10.1007/s11033-022-08015-7](https://doi.org/10.1007/s11033-022-08015-7)
25. Filonzi L, Ardenghi A, Rontani PM, et al. Molecular Barcoding: A Tool to Guarantee Correct Seafood Labelling and Quality and Preserve the Conservation of Endangered Species. *Foods*. 2023;12(12). doi:[10.3390/foods12122420](https://doi.org/10.3390/foods12122420)
26. Hebert PD, Cywinska A, Ball SL, deWaard JR. Biological identifications through DNA barcodes. *Proc Biol Sci*. 2003;270(1512):313-321. doi:[10.1098/rspb.2002.2218](https://doi.org/10.1098/rspb.2002.2218)
27. Dawnay N, Ogden R, McEwing R, Carvalho GR, Thorpe RS. Validation of the barcoding gene COI for use in forensic genetic species identification. *Forensic Science International*. 2007;173(1):1-6. doi:[10.1016/j.forsciint.2006.09.013](https://doi.org/10.1016/j.forsciint.2006.09.013)
28. Shokoohmand M, Zolgharnein H, Mashjoor S, Laloi F, Foroughmand AM, Savari A. Analysis of Genetic Diversity of White Shrimp (*Metapenaeus affinis*) from the Northwest of the Persian Gulf Using Microsatellite Markers. *Turkish Journal of Fisheries and Aquatic Sciences*. 2018;18:385-394. doi:[10.4194/1303-2712-v18_3_04](https://doi.org/10.4194/1303-2712-v18_3_04)
29. Zhang D, Liu T, Gong F, Jiang S. Shotgun assembly of the first mitochondrial genome of *Metapenaeus* (*Metapenaeus ensis*) with phylogenetic consideration. *Mitochondrial DNA A DNA Mapp Seq Anal*. 2016;27(5):3689-3690. doi:[10.3109/19401736.2015.1079857](https://doi.org/10.3109/19401736.2015.1079857)
30. Shringarpure S, Xing EP. Effects of sample selection bias on the accuracy of population structure and ancestry inference. *G3 (Bethesda)*. 2014;4(5):901-911. doi:[10.1534/g3.113.007633](https://doi.org/10.1534/g3.113.007633)
31. Prasertlux S, Khamnamtong B, Wisuntorn E, et al. Genetic diversity and population differentiation of wild and domesticated banana shrimp *Fenneropenaeus merguensis*: Applications for development of its breeding program. *Regional Studies in Marine Science*. 2024;69:103309. doi:[10.1016/j.rsma.2023.103309](https://doi.org/10.1016/j.rsma.2023.103309)
32. My TTA, Dat ND, Hung NQ. Occurrence and Characteristics of Microplastics in Wild and Farmed Shrimps Collected from Cau Hai Lagoon, Central Vietnam. *Molecules*. 2023;28(12). doi:[10.3390/molecules28124634](https://doi.org/10.3390/molecules28124634)
33. Thanh TD. Tam Giang – Cau Hai: lagoon resources potential and orientation for management. *Vietnam Journal of Marine Science and Technology*. 2007;7:53-62.
34. Boonstra WJ, Hanh TTH. Adaptation to climate change as social–ecological trap: a case study of fishing and aquaculture in the Tam Giang Lagoon, Vietnam. *Environment, Development and Sustainability*. 2015;17(6):1527-1544. doi:[10.1007/s10668-014-9612-z](https://doi.org/10.1007/s10668-014-9612-z)
35. Trang CTT, Thanh T, Thanh TD, Vinh VD, Tu TA. Assessment of the environmental carrying capacity of pollutants in Tam Giang-Cau Hai Lagoon (Viet Nam) and solutions for the environment protection of the lagoon. *Science of The Total Environment*. 2021;762:143130. doi:[10.1016/j.scitotenv.2020.143130](https://doi.org/10.1016/j.scitotenv.2020.143130)
36. Luong LT, Thang LT. Study on histological structure of greasyback shrimp (*Metapenaeus ensis* De Haan, 1850) male gonad in Tam Giang-Cau Hai lagoon, Thua Thien Hue. *Journal of Fisheries Science and Technology*. 2019;03:83-90.
37. Weight S. Evolution and the genetics of population. In: Wright S, ed. *Variability Within and Among Natural Populations*. University of Chicago Press; 1978:10-15.