The Open Access Israeli Journal of Aquaculture – Bamidgeh

As from **January 2010** The Israeli Journal of Aquaculture - Bamidgeh (IJA) will be published exclusively as **an on-line Open Access (OA)** quarterly accessible by all AquacultureHub (<u>http://www.aquaculturehub.org</u>) members and registered individuals and institutions. Please visit our website (<u>http://siamb.org.il</u>) for free registration form, further information and instructions.

This transformation from a subscription printed version to an on-line OA journal, aims at supporting the concept that scientific peer-reviewed publications should be made available to all, including those with limited resources. The OA IJA does not enforce author or subscription fees and will endeavor to obtain alternative sources of income to support this policy for as long as possible.

Editor-in-Chief

Dan Mires

Editorial Board

| Sheenan Harpaz | Agricultural Research Organization Beit Dagan, Israel |
|------------------|---|
| Zvi Yaron | Dept. of Zoology Tel Aviv University Tel Aviv, Israel |
| Angelo Colorni | National Center for Mariculture, IOLR Eilat, Israel |
| Rina Chakrabarti | Aqua Research Lab Dept. of Zoology University of Delhi |
| Ingrid Lupatsch | Swansea University Singleton Park, Swansea, UK |
| Jaap van Rijn | The Hebrew University Faculty of Agriculture Israel |
| Spencer Malecha | Dept. of Human Nutrition, Food and Animal Sciences University of Hawaii |
| Daniel Golani | The Hebrew University of Jerusalem Jerusalem, Israel |
| Emilio Tibaldi | Udine University Udine, Italy |

Published under auspices of **The Society of Israeli Aquaculture and Marine Biotechnology (SIAMB), University of Hawaii at Manoa Library** and **University of Hawaii Aquaculture Program** in association with **AquacultureHub** http://www.aquaculturehub.org





AquacultureHub

ISSN 0792 - 156X

 $\ensuremath{\textcircled{C}}$ Israeli Journal of Aquaculture - BAMIGDEH.

PUBLISHER: Israeli Journal of Aquaculture - BAMIGDEH -Kibbutz Ein Hamifratz, Mobile Post 25210, ISRAEL Phone: + 972 52 3965809 <u>http://siamb.org.il</u>

Copy Editor Ellen Rosenberg

Genetic Diversity of Gilthead Seabream (Sparus aurata) Broodstocks as Determined by RAPD-PCR

Guldehen Bilgen¹, Suleyman Akhan^{2*}, Muhammed Arabaci³, and Ismail Oguz¹

¹ Department of Animal Science, Faculty of Agriculture, Ege University, 35100 Bornova, Izmir, Turkey

² Department of Aquaculture, Faculty of Fisheries, Rize University, 53100 Rize, Turkey

³ Department of Fisheries, Faculty of Agriculture, Yuzuncu Yil University, 65080 Van, Turkey

(Received 17.3.07, Accepted 3.5.07)

Key words: RAPD-PCR, gilthead seabream, Sparus aurata, broodstock, genetic diversity

Abstract

The objective of this study was to determine the genetic diversity in gilthead seabream broodstocks from two hatcheries on the northern Aegean Coast of Turkey by RAPD-PCR. Forty primers were tested for each broodstock. Twenty-five produced scorable RAPD bands in stock from the Seferihisar hatchery and 28 in stock from the Aliaga hatchery. Nineteen revealed patterns with scorable amplified primers in both broodstocks. Depending on primer, the number of bands varied 3-16, ranging in size from 438 to 2520 base pairs (bp). The average genetic similarity within stocks was 0.466 for the Seferihisar stock and 0.617 for the Aliaga. The average genetic similarity between the two broodstocks was 0.420, lower than the values within the broodstocks, and the genetic distance between the two broodstocks was 0.245.

Introduction

The gilthead seabream (*Sparus aurata*) inhabits the Mediterranean and Atlantic coasts of Europe and is one of the most important cultured marine fish in the Mediterranean and Adriatic Seas. The commercial importance of gilthead seabream has generated intense interest in its molecular genetics. Funkenstein et al. (1990) first reported on mitochondrial DNA polymorphism of gilthead seabream broodstocks in Israel by using RFLP analysis of the whole mitochondrial DNA (mtDNA) molecule. Magoulas et al. (1995) reported on mtDNA polymorphism for

Greece broodstocks. Palma et al. (2001) studied developmental stability and genetic heterozygosity in wild and cultured gilthead seabream stocks. Alarcon et al. (2004) compared cultivated and wild seabream stocks from the Atlantic and Mediterranean coasts using allozymes, microsatellite, and mtDNA markers. De Innocentiis et al. (2004) investigated the genetic variability of gilthead seabream populations from the Atlantic Ocean, Mediterranean Sea, and Adriatic Sea using microsatellite markers.

In Turkey, mariculture of seabream has

^{*} Corresponding author. E-mail: akhan@ktu.edu.tr

Bilgen et al.

continuously grown since it began in the late 1980s. The aim of the present study was to determine the genetic diversity within and between two Turkish gilthead seabream broodstocks using random amplified polymorphic DNA-polymerase chain reaction (RAPD-PCR), a relatively simple, quick, and inexpensive technique that requires no target DNA sequence information.

Materials and Methods

Fish samples. Gilthead seabream (4 years old) were provided by two private hatcheries in Seferihisar and Aliaga, in Izmir province, western Turkey. The Seferihisar broodstock originated from Sigacik Bay and the Bostanli coast of Izmir Gulf (Fig. 1). The Aliaga broodstock originated from the Karatas lagoon in the Adana province of eastern Mediterranean. All broodstock were captured as juveniles and reared in cage farms until market size. Then

they were transferred to hatcheries and used for seed production. Fifteen randomly chosen fish from each hatchery were used in this study.

DNA extraction. Blood samples (0.5-1 ml) were collected from the caudal vein using a 5-ml EDTA-coated syringe. Samples were put into separate 5-ml tubes containing K₃-EDTA. DNA was isolated from the samples according to Dunnington et al. (1990). The DNA pellet was dissolved in 100 μ l of TE buffer (10mM Tris, 1mM EDTA, pH 8) and the concentration of each DNA sample was checked by spectrophotometer.

RAPD-PCR. Forty 10-mer random oligonucleotide primers (OPA and OPB, each series consisting of 20 primers; Operon Technologies, Alameda, CA, USA) were used to amplify the genomic DNA. Amplification reactions were carried out on a final volume of 15 μ l containing 25 ng DNA, 100 μ M each of



Fig. 1. Hatcheries and sampling regions. \bullet = hatcheries; \blacktriangle = origins of Seferihisar broodstock: BS = Bostanli coast, SB = Sigacik Bay; \blacktriangledown origin of Aliaga broodstock: KL = Karatas lagoon.

dATP, dTTP, dCTP, and dGTP (Boehringer Mannheim, Germany), 15 ng of the primer, 1x Super Therm polymerase buffer (20 mM Tris-HCI, pH 8, 100 mM NaCl, 0.1 mM EDTA, 1 mM DTT, 50% glycerol), and 1 unit of Super Therm DNA polymerase enzyme (SR Products, UK). The PCR reactions were performed in a thermal cycler (Appligene) programmed for initial denaturation in 30 s at 94°C followed by 35 cycles of 25 s at 94°C, 45 s at 35°C, and 1 min at 72°C. Amplification was terminated by a final extension step of 5 min at 72°C. Amplification products were electrophoresed in 1.5% agarose gel, viewed on a UV transilluminator after staining with ethidium bromide (2 µg/ml), and photographed. Lambda DNA-EcoR I/Hind III double digest was used as a molecular size marker and run parallel to the amplified products.

Data analysis. RAPD bands were scored using a data matrix for the presence (1) or absence (0) of markers for the 19 primers that produced polymorphic bands in both broodstocks. Sizes in base pairs (bp) were inferred by comparison with lambda DNA-EcoR I/Hind III. The RAPD patterns of individuals were compared within and between broodstocks. A RAPD data matrix was used to determine the genetic similarity between individuals in terms of average band sharing frequencies (Nei and Li, 1979). The genetic similarity index (band sharing) was calculated as: $S_{xy} = 2(N_{xy})/(N_x +$ N_y), where N_{xy} is the number of bands present in both individuals, N_x is the number of bands in individual x, and N_v is the number of bands in individual y.

The Student's *t* test (Steel and Torrie, 1980) was performed to compare average genetic similarity between Seferihisar and Aliaga broodstocks.

Genetic distance was calculated as described by Lynch (1991), $D_{ij} = -\ln S_{ij}/(\sqrt{S_i \times S_j})$, where D_{ij} is the approximate genetic distance between broodstocks i and j, S_{ij} is the average genetic similarity index between broodstocks i and j, S_i is the average similarity index in broodstock i, and S_j is the average similarity index in broodstock j. A dendrogram was constructed using Ward's method of hierarchical cluster analysis of JMP (SAS, 1996)

Results

Of the 40 primers, 25 revealed a pattern with scorable amplified primers in the Seferihisar broodstock and 28 revealed such a pattern in the Aliaga broodstock. Nineteen revealed patterns with scorable amplified primers in the both broodstocks (Table 1). Primers OPA-7, OPA-9, OPA-10, OPA-11, OPA-16, OPA-18, OPB-1, OPB-2, OPB-5, OPB-8, OPB-11, OPB-12, OPB-17, and OPB-18 exhibited the highest quality band patterns.

Genetic similarity averaged 0.466 within the Seferihisar broodstock (range 0.273-0.777; Table 2) and 0.617 within the Aliaga broodstock (range 0.420-0.770; Table 3), a significant difference of $p \le 0.001$. The genetic similarity between Aliaga and Seferihisar broodstock averaged 0.420 (range 0.197-0.712; Table 4). The genetic distance between the two broodstocks was 0.245.

The dendrogram (Fig. 2) resulted in two main clusters: (a) nine fish from the Seferihisar stock (S1-3, S5-7, S9, S10, S15) and (b) all the Aliaga stock (A1-A15) plus six fish from the Seferihisar stock (S4, S8, S11-14).

Discussion

Seabream is one of the most important cultured fish species in the Mediterranean. Efforts to genetically improve its production traits (Knibb et al., 1997) may decrease genetic variation in cultured fish stocks as compared to wild stocks (Youngson et al., 2001). Palma et al. (2001), who studied the difference between cultivated and wild stocks, reported that rare alleles in the cultivated stocks disappeared as a result, in their opinion, of breeding techniques that rely on the same parental stock for several generations.

In our study, the average genetic similarity between the two stocks was 0.420, lower than the average within either stock. Bielawski and Puma (1997) used RAPD to detect genetic similarity within and between Atlantic coast striped bass (*Morone saxatilis*) and also found that the interpopulation average similarity (88.4%) was lower than the intrapopulation average similarity (93.8%). De Innocentiis et al. (2004) reported high polymorphism (7-38 alleBilgen et al.

| Primer | Sequence (5' – 3') | Sef | erihisar | A | liaga |
|--------|--------------------|----------------|-----------------------|----------------|-----------------------|
| | | Bands (no.) | Base pairs (range) | Bands (no.) | Base pairs (range) |
| OPA-4 | AATCGGGCTG | 8 | 480-1433 | 7 | 512-1716 |
| OPA-7 | GAAACGGGTG | 16 | 438-2283 | 12 | 438-2283 |
| OPA-9 | GGGTAACGCC | 9 | 510-1270 | 8 | 510-1270 |
| OPA-10 | GTGATCGCAG | 3 | 462-594 | 9 | 462-1032 |
| OPA-11 | CAATCGCCGT | 12 | 461-1567 | 10 | 491-1567 |
| OPA-13 | CAGCACCCAC | 3 | 590-804 | 5 | 590-1318 |
| OPA-14 | TCTGTGCTGG | 5 | 555-944 | 4 | 587-944 |
| OPA-18 | AGGTGACCGT | 11 | 497-1076 | 7 | 497-978 |
| OPA-19 | CAAACGTCGG | 4 | 612-1345 | 12 | 506-1451 |
| OPA-20 | GTTGCGATCC | 13 | 482-1592 | 13 | 482-1592 |
| OPB-1 | GTTTCGCTCC | 15 | 508-1936 | 15 | 465-1936 |
| OPB-5 | TGCGCCCTTC | 11 | 619-1599 | 16 | 459-1599 |
| OPB-6 | TGCTCTGCCC | 9 | 494-922 | 10 | 494-942 |
| OPB-7 | GGTGACGCAG | 11 | 483-1362 | 11 | 483-1362 |
| OPB-8 | GTCCACACGG | 10 | 468-1254 | 9 | 468-1254 |
| OPB-11 | GTAGACCCGT | 15 | 523-2520 | 15 | 523-2520 |
| OPB-12 | CCTTGACGCA | 10 | 453-1029 | 13 | 453-1811 |
| OPB-17 | AGGGAACGAG | 12 | 456-1640 | 11 | 533-1640 |
| OPB-18 | CCACAGCAGT | 8 | 583-1310 | 7 | 583-1310 |
| Total | - | 185 | - | 194 | - |

Table 1. RAPD profiles of Seferihisar and Aliaga seabream broodstocks (19 primers).

les/locus) for four microsatellite loci among gilthead seabream populations from the Atlantic Ocean, Mediterranean Sea, and Adriatic Sea, concluding that the Mediterranean population differed from the Atlantic and Adriatic populations.

Genetic diversity within the Seferihisar broodstock was greater than within the Aliaga broodstock. Seferihisar broodstock originated from two areas, which may explain the greater variation in Seferihisar broods.

The dendrogram shows no strong genetic differentiation between the two stocks as some of the Seferihisar specimens fell into the same cluster as the Aliaga broodstock. This suggests some genetic exchange between the Aegean Sea and eastern Mediterranean wild seabream populations.

The level of genetic diversity is an important consideration in establishing a broodstock for selective breeding because it provides an indication of the scope for selective progress (Silverstein et al., 2001). In this respect, because of its relatively greater genetic diversity, the Seferihisar broodstock could serve as material for selective breeding.

In conclusion, RAPD effectively determined the genetic diversity within and between two gilthead seabream broodstocks in Turkey. In this study, we worked on limited strains. Future studies should examine wild as well as cultivated strains.

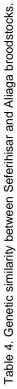
| | S1 | S2 | S3 | S4 | S5 | S6 | S7 | S8 | S9 | S10 | S11 | S12 | S13 | S14 |
|----------|---|-------------|--------------|------------|----------|-------|-------|-------|-----------|-------|-------|-------|-------|-------|
| S2 S3 | 0.411 0.391 | 0.541 | | | | | | | | | | | | |
| S4 8 | 0.352 | 0.524 | 0.638 | | | | | | | | | | | |
| S5 | 0.327 | 0.342 | 0.457 | 0.423 | | | | | | | | | | |
| S6 | 0.396 | 0.273 | 0.324 | 0.389 | 0.374 | | | | | | | | | |
| S7 | 0.387 | 0.516 | 0.486 | 0.450 | 0.364 | 0.401 | | | | | | | | |
| S8 | 0.460 | 0.525 | 0.564 | 0.780 | 0.375 | 0.351 | 0.532 | | | | | | | |
| S9 | 0.494 | 0.327 | 0.398 | 0.383 | 0.371 | 0.403 | 0.490 | 0.482 | | | | | | |
| S10 | 0.398 | 0.367 | 0.515 | 0.376 | 0.337 | 0.287 | 0.460 | 0.460 | 0.518 | | | | | |
| S11 | 0.387 | 0.464 | 0.451 | 0.590 | 0.325 | 0.265 | 0.521 | 0.659 | 0.455 | 0.380 | | | | |
| S12 | 0.460 | 0.515 | 0.584 | 0.777 | 0.384 | 0.378 | 0.499 | 0.764 | 0.450 | 0.385 | 0.667 | | | |
| S13 | 0.451 | 0.483 | 0.541 | 0.674 | 0.387 | 0.412 | 0.477 | 0.689 | 0.398 | 0.442 | 0.562 | 0.760 | | |
| S14 | 0.401 | 0.427 | 0.641 | 0.720 | 0.381 | 0.357 | 0.428 | 0.718 | 0.390 | 0.482 | 0.563 | 0.750 | 0.672 | |
| S15 | 0.404 | 0.396 | 0.590 | 0.421 | 0.356 | 0.246 | 0.390 | 0.470 | 0.381 | 0.523 | 0.392 | 0.447 | 0.523 | 0.499 |
| Tat | Table 3. Genetic similarity within Aliaga broodstock. | etic simile | arity withir | ר Aliaga t | roodstoc | ÷ | | | | | | | | |
| | A1 | A2 | A3 | A4 | A5 | AG | A7 | A8 | <i>A9</i> | A10 | A11 | A12 | A13 | A14 |
| A2 | 0.588 | | | | | | | | | | | | | |
| A3 | 0.536 | 0.631 | | | | | | | | | | | | |
| A4 | 0.595 | 0.634 | 0.655 | | | | | | | | | | | |
| A5 | 0.546 | 0.561 | 0.604 | 0.627 | | | | | | | | | | |
| A6 | 0.497 | 0.552 | 0.694 | 0.642 | 0.636 | | | | | | | | | |
| A7 | 0.553 | 0.594 | 0.577 | 0.670 | 0.628 | 0.680 | | | | | | | | |
| A8 | 0.587 | 0.665 | 0.701 | 0.660 | 0.711 | 0.728 | 0.670 | | | | | | | |
| A9 | 0.516 | 0.485 | 0.566 | 0.575 | 0.586 | 0.637 | 0.661 | 0.663 | | | | | | |
| A10 | 0.612 | 0.642 | 0.696 | 0.694 | 0.694 | 0.736 | 0.706 | 0.793 | 0.655 | | | | | |
| A11 | 0.539 | 0.517 | 0.627 | 0.554 | 0.551 | 0.558 | 0.464 | 0.550 | 0.420 | 0.590 | | | | |
| A12 | 0.504 | 0.581 | 0.635 | 0.675 | 0.680 | 0.701 | 0.730 | 0.770 | 0.693 | 0.755 | 0.517 | | | |
| A13 | 0.504 | 0.565 | 0.666 | 0.716 | 0.627 | 0.663 | 0.656 | 0.674 | 0.622 | 0.737 | 0.555 | 0.737 | | |
| A14 | 0.507 | 0.550 | 0.644 | 0.640 | 0.583 | 0.625 | 0.612 | 0.676 | 0.560 | 0.735 | 0.515 | 0.711 | 0.751 | |
| A15 | 0.430 | 0.519 | 0.592 | 0.569 | 0.515 | 0.606 | 0.589 | 0.627 | 0.585 | 0.636 | 0.548 | 0.658 | 0.601 | 0.610 |

Table 2. Genetic similarity within Seferihisar broodstock.

Genetic diversity of gilthead seabream by RAPD-PCR

221

| S | 7 | Ñ | S3 | S4 | S5 | S6 | S7 | S8 | S9 | S10 | S11 | S12 | S13 | S14 | S15 |
|---------|-----------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| 0.2 | 0.287 0.3 | 0.318 | 0.416 | 0.454 | 0.257 | 0.324 | 0.380 | 0.471 | 0.326 | 0.332 | 0.363 | 0.466 | 0.442 | 0.436 | 0.345 |
| ю. О | 04 0.3 | 44 | 0.377 | 0.457 | 0.236 | 0.235 | 0.331 | 0.559 | 0.354 | 0.326 | 0.422 | 0.520 | 0.463 | 0.485 | 0.361 |
| 0.3 | 62 0.3 | 369 | 0.406 | 0.524 | 0.317 | 0.251 | 0.343 | 0.527 | 0.407 | 0.293 | 0.443 | 0.592 | 0.472 | 0.493 | 0.368 |
| 0.2 | 86 0.3 | 120 | 0.415 | 0.559 | 0.326 | 0.268 | 0.352 | 0.549 | 0.332 | 0.300 | 0.439 | 0.591 | 0.494 | 0.530 | 0.312 |
| 0.3 | 67 0.4 | 601 | 0.466 | 0.582 | 0.318 | 0.299 | 0.443 | 0.538 | 0.414 | 0.304 | 0.476 | 0.634 | 0.495 | 0.526 | 0.299 |
| 0. 3 | 57 0.4 | 124 | 0.474 | 0.587 | 0.329 | 0.307 | 0.362 | 0.532 | 0.358 | 0.286 | 0.417 | 0.620 | 0.482 | 0.531 | 0.327 |
| 0.2 | 99 0.3 | 376 | 0.436 | 0.556 | 0.310 | 0.269 | 0.351 | 0.544 | 0.262 | 0.274 | 0.387 | 0.576 | 0.509 | 0.487 | 0.278 |
| ю. О | 64 0.4 | 140 | 0.494 | 0.612 | 0.361 | 0.319 | 0.399 | 0.592 | 0.379 | 0.345 | 0.491 | 0.688 | 0.560 | 0.587 | 0.462 |
| ю. О | 54 0.4 | 51 | 0.529 | 0.561 | 0.349 | 0.290 | 0.416 | 0.528 | 0.329 | 0.310 | 0.440 | 0.616 | 0.529 | 0.496 | 0.375 |
| ю. Э | 61 0.4 | 126 | 0.511 | 0.683 | 0.372 | 0.331 | 0.426 | 0.623 | 0.382 | 0.357 | 0.510 | 0.712 | 0.599 | 0.609 | 0.415 |
| 0.3 | 27 0.2 | 83 | 0.351 | 0.408 | 0.228 | 0.197 | 0.295 | 0.437 | 0.325 | 0.254 | 0.329 | 0.443 | 0.393 | 0.414 | 0.293 |
| ю. О | 80 0.4 | 137 | 0.491 | 0.644 | 0.386 | 0.346 | 0.375 | 0.626 | 0.377 | 0.334 | 0.519 | 0.686 | 0.631 | 0.590 | 0.361 |
| ю. Э | 03 0.4 | t07 | 0.507 | 0.600 | 0.364 | 0.226 | 0.341 | 0.579 | 0.356 | 0.340 | 0.491 | 0.646 | 0.530 | 0.605 | 0.363 |
| 0.2 | 66 0.3 | 399 | 0.449 | 0.634 | 0.305 | 0.252 | 0.327 | 0.591 | 0.317 | 0.317 | 0.450 | 0.627 | 0.559 | 0.567 | 0.340 |
| Ю. Э | 02 0.3 | 358 | 0.418 | 0.539 | 0.308 | 0.226 | 0.333 | 0.487 | 0.297 | 0.259 | 0.429 | 0.541 | 0.513 | 0.509 | 0.333 |



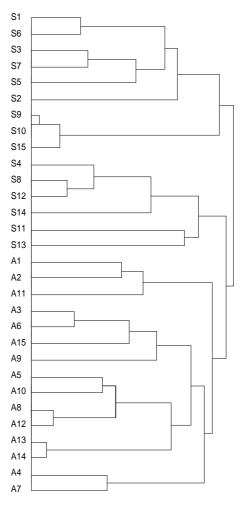


Fig. 2. Genetic relationship between Seferihisar and Aliaga broodstocks.

Acknowledgements

This study was supported by TUBITAK (The Scientific and Technical Research Council of Turkey) project no. VHAG-1673. We would like to thank TUBITAK and the managers of the two private hatcheries for their support and cooperation.

References

Alarcon J.A., Magoulas A., Georgakopoulos T., Zouros E. and M.C. Alvarez, 2004. Genetic comparison of wild and cultivated

222

Bilgen et al.

European populations of the gilthead seabream (*Sparus aurata*). *Aquaculture*, 230: 65-80.

Bielawski J.P. and D.E. Pumo, 1997. Randomly amplified polymorphic DNA (RAPD) analysis of Atlantic coast striped bass. *Heredity*, 78:32-40.

De Innocentiis S., Lesti A., Livi S., Rossi A.R., Crosetti D. and L. Sola, 2004. Microsatallite markers reveal population structure in gilthead seabream *Sparus auratus* from the Atlantic Ocean and Mediterranean Sea. *Fish. Sci.*, 70:852-859.

Dunnington E.A., Gal O., Plotsky Y., Haberfeld A., Kirk T., Golberg A., Lavi U., Cahaner A., Siegel P.B. and J. Hillel, 1990. DNA fingerprints of chickens selected for high and low body weight for 31 generations. *Anim. Genet.*, 21:221-231.

Funkenstein B., Cavari B., Stadie T. and E. Davidovitch, 1990. Restriction site polymorphism of mitochondrial DNA of the gilthead seabream (*Sparus aurata*) broodstock in Eilat, Israel. *Aquaculture*, 89:217-223

Knibb W., Gorshkova G. and S. Gorshkov, 1997. Selection for growth in the gilthead seabream, *Sparus aurata* L.. *Israeli J. Aquac.* – *Bamidgeh*, 49:57-66.

Lynch M., 1991. Analysis of population genetic structure by DNA fingerprinting. pp. 112-126. In: I. Burke, G. Dolf, A.J. Jeffreys, R. Wolff (eds.). *DNA Fingerprints Approaches and Application*. Birkhauser Verlag AG, Basel, Switzerland.

Magoulas A., Sophronides K., Patarnello T., Hatzilaris E. and E. Zouros, 1995. Mitochondrial DNA variation in an experimental stock of gilthead seabream (*Sparus aurata*). *Mol. Mar. Biol. Biotech.*, 4:110-116.

Nei M. and W.H. Li, 1979. Mathematical modeling for studying genetic variation in terms of restriction endonucleases. *Proc. Natl. Acad. Sci.*, 76:5269-5273.

Palma J., Alarcon J.A., Alvarez C., Zouros E., Magoulas A. and J.P. Andrade, 2001. Developmental stability and genetic heterozy-gosity in wild and cultured stocks of gilthead seabream (*Sparus aurata*). J. Mar. Biol. Assoc. UK., 81:283-288.

SAS, 1996. JMP User's Guide, Version 3.2.2, SAS Institute Inc. Cary, NC.

Silverstein J.T., Rexroad III C.E. and T.L. King, 2004. Genetic variation measured by microsatellites among three strains of domesticated rainbow trout (*Oncorhynchus mykiss*, Walbaum. *Aquac. Res.*, 35:40-48.

Steel R.G.D. and J.H. Torrie, 1980. *Principles and Procedures of Statistics: A Biometrical Approach*, 2nd ed. McGraw-Hill, New York. 633 pp.

Youngson A.F., Dosdat A., Saroglia M. and W.C. Jordan, 2001. Genetic interaction between marine finfish species in European aquaculture and wild conspecifics. *J. Appl. lchthyol.*, 17:153-162.