

The IJA appears exclusively as a peer-reviewed on-line open access journal at <u>http://www.siamb.org.il</u> Sale of IJA papers is strictly forbidden.



Offshore Farming of the Mediterranean Amberjack (Seriola dumerili) in the Northeastern Mediterranean

Erdal Yilmaz* and Hülya Şereflişan

Faculty of Fisheries and Aquaculture, Mustafa Kemal University, Kapalı Yüzme Havuzu Yanı, İskenderun Hatay, Turkey

(Received 4.5.10, Accepted 31.7.10)

Key words: Seriola dumerili, offshore, farming, feeding, Mediterranean

Abstract

The growth rate, survival, and feeding parameters of Mediterranean amberjack (Seriola dumerili Risso, 1810) cultivated in offshore cages in the Gulf of İskenderun (Hatay, Turkey) were investigated from July 2006 to March 2008. Two offshore cages (339 m^3) were placed at a depth of 10 m in a sheltered area 500 m off the coast. Juveniles (101.97±14.54 g), caught with seine nets 400-1200 m from the coast, were stocked in the cages at 250 per cage. Fish in one cage, Group A, were fed commercial pellets for all 20 months whilst fish in the second cage, Group B, were fed pellets for the first 16 months and frozen sardines for the following four. The mean live weight, survival, feed conversion ratio, daily feed consumption, and water quality parameters were recorded monthly. Group B reached a significantly (p < 0.05) greater weight (2785.00±64.91 g) than Group A (2441.14±38.13 g). Food conversion ratios ranged 1.16-2.46 for group A and 1.10-2.28 for group B, low when compared with other studies on amberjacks in the Mediterranean. Average mortality was 25%, due to a *Zeuxapta seriolae* (Monogenea: Heteraxinidae) infection in December 2006 (20%) and fish handling for measurement in June 2007 (~5%). The farmed amberjacks had significantly higher protein and lipid contents than their wild counterparts at the end of the 20-month feeding period. Amberjacks found frozen sardines more appealing than pellets, especially during the winter. In conclusion, it is advised to feed Mediterranean amberjack frozen sardines or other trash fish during the winter grow-out period. Their rapid growth rate, as well as their adaptability to farming, make offshore net cages a promising system for the aquaculture of this species.

Yilmaz and Şereflişan

Introduction

The Mediterranean amberjack, *Seriola dumerili* (Risso 1810), also referred to as the greater amberjack, is a carnivorous pelagic fish (Carangidae) that satisfies the criteria for selection of new aquaculture species (Marino et al., 1995), i.e., it has a rapid growth rate (Andaloro et al., 1992), easily adjusts to captivity, has excellent flesh quality, and is sold for a high market price (Nakada, 2000). It is distributed throughout tropical and subtropical regions except the Pacific Ocean (Laroche et al., 1984). In the eastern Atlantic, this species is distributed from the Bay of Biscay to South Africa, whereas in the western Atlantic, it inhabits the area from Nova Scotia to Brazil, including the Gulf of Mexico, the Caribbean, and Bermuda (Fischer, 1978). Amberjack is abundant in the Mediterranean area, except in the Black and north Adriatic Seas, (Andaloro et al., 1992).

The morphological and ecological characteristics of *S. dumerili* are similar to *S. quinqueradiata* which has been widely reared in Japan for over 25 years. *Seriola dumerili* is also reared in Japan and is imported from China to Japan. It has a high growth rate in the natural environment (Thompson et al., 1999) and in culture conditions (Jover et al., 1999; Mazzola et al., 2000). Hence, it has become one of the most important aquaculture species in the Mediterranean region as well as in Japan (Nakata, 2000; Takakuwa et al., 2006). It adapts easily to a captive environment but reproduction in culture conditions is difficult, although it has recently been successfully reproduced in Greece and Malta and Japanese scientists have successfully reared larvae since 1987 (Tachihara et al., 1993).

The commercial culture of *S. dumerili* in the Mediterranean is based on capture of wild juveniles that are fattened to marketable size (Lazzari et al., 2000; Pastor et al., 2000). On-growing in Mediterranean countries (Spain, Italy) as well as Japan generally occurs in net cages, using raw, fresh, frozen, or chopped fish (*Sardinella aurita, Boops boops, Trachurus trachurus, Spicara maena*). In the Mediterranean, juveniles have been reared in concrete tanks (Cavaliere et al., 1989; Garcia et al., 1993) and inshore net cages (Porrello et al., 1993). This investigation presents results of the cultivation of amberjack in offshore cages using simple low-cost installation techniques. Growth rate, survival, and feeding parameters of two groups of fish fed different diets in the northeastern Mediterranean were tested.

Materials and Methods

Feeding trials were carried out between July 2006 and April 2008 in the Gulf of İskenderun (Hatay, Turkey) in a sheltered area 500 m off the coast. Two offshore cages of 339 m³ (12 m diameter \times 3 m, 12 and 18-mm mesh for juveniles and adults, respectively) were moored onto artificial reefs at a depth of 10 m. Together with local fishermen, the research team caught juveniles (101.97±14.54 g) during the summer of 2006 using seine nets with 14-16 mm mesh, 400-1200 m from the coast. The fish were weighed and placed in a 2-m³ PVC tank on the fishing boat. The tank was equipped with a flow-through seawater system during transport to the cages. Two hundred and fifty fish were stocked into each cage as soon as possible.

The juveniles were fed three diets composed of frozen sardines (*Sardina pilchardus*) and pellets at rates of 75/25, 50/50, and 25/75 for the first, second, and third months (July, August, and September 2006, respectively) to wean them to the dry commercial diet. Frozen sardines were initially offered in small minced pieces (heads removed) but, as the juveniles grew, the sardines were chopped to adequate sizes. After weaning, fish in one cage, Group A, were fed commercial pellets for 17 months and fish in the second cage, Group B, were hand-fed commercial pellets for 13 months (from November 2006 to to December 2007) and frozen sardines for the next 4 months. Fish were fed to satiation three times a day throughout the experimental period. The daily amount of food administered was recorded for both groups throughout the experimental period.

The mean body weight, specific growth rate, survival, feed conversion ratio, and total amount of food administered (as a function of body weight, %) were calculated on a monthly basis. Growth performance data were analyzed as follows: daily specific growth rate (%/day) in somatic live weight as SGR =100 $[(W_t/W_0)^{1/t} - 1]$, where W_t = weight on

day t, W_0 = weight on day 0, and t = time elapsed in days; and the feed conversion ratio (FCR) as the amount of food consumed (dry weight in g) divided by the growth in biomass (wet weight in g) for a specific time period. Water quality parameters were measured on a daily basis, dissolved oxygen and temperature with a YSI 52 model oxygen meter (Yellow Springs Instruments Co., Ohio, USA), pH with an ORION 420A model, and salinity with an ATAGO hand refractometer.

At the end of the study, standard laboratory methods (AOAC, 1990) were used to determine the proximate compositions of fillets of the cultured and wild amberjacks as follows: moisture was determined by oven-drying at 105°C for 24 h, crude protein (N x 6.25) by the Kjeldahl method, and crude ash by combustion in a muffle furnace at 550°C for 16 h. Total lipid concentration was determined by the extract with chloroform-methanol method (Bligh and Dyer, 1959). Differences among groups in growth performance and proximate fillet composition were tested with Kruskal-Wallis one-way analysis of variance by ranks (SPSS 10.0 for Windows) followed by the Duncan non-parametric multiple comparison procedure (Zar, 1984).

Results

Monthly temperature, salinity, dissolved oxygen, and pH are shown Fig. 1. Water temperature at the surface ranged 16.5-19.0°C in December-February, 17.0-20.0°C in March-May, 24.5-29.0°C in June-August, and 24.0-28.0°C in September-November. For the same months, salinity was 35.5-36.1, 36.4-37.4, 37.3-37.8, and 36.2-37.1, dissolved oxygen 8.4-8.7, 7.7-8.1, 6.7-7.2, and 6.7-7.9, and pH 8.4-8.7, 8.4-8.5, 8.6-8.7, and 8.0-8.3.



Fig. 1. Monthly salinity, pH, dissolved oxygen, and temperature in offshore cages used to cultivate the Mediterranean amberjack (*Seriola dumerili*).



Fig. 2. Monthly average live weight of *Seriola dumerili* fed pellets only (Group A) or pellets and sardines (Group B).

The wild Mediterranean amberjack juveniles adapted well to captivity conditions and accepted the food provided from their arrival the offshore culture system in onwards. They had a high growth rate throughout the 20 months of rearing. Fish in Group В (2785.00±64.91 g) reached a higher weight (p < 0.05) than those in Group A $(2441\pm38.13 \text{ g})$ by the end of the study (Fig. 2).

At the end of the trial, mortality was 24.8% and 23.2% in Groups A and B, respectively, due to infection (20%) by *Zeuxapta seriolae* (Monogenea: Heteraxinidae) in December 2006 and handling for monthly measurements (~5%) in June 2007 (Table 1).

Survival and feeding parameters did not differ significantly between groups. There were no significant differences in daily live weight gain or SGR except at the end of the experiment (March 2008). The feeding rates were constant, except when they dropped in September 2006. SGR was lowest in December 2006 due to the low water temperature and *Z. seriolae* infection (Fig. 3). FCR did not significantly differ and decreased with the increasing water temperature, as expected (Fig. 4).

Table 1. Average weight, survival, and feeding parameters of Mediterranean amberjack fed a commercial feed for 20 months (Group A) or a commercial feed for 16 months and frozen sardines for the remaining four months (Group B).

Month	Mean wt±SD		Wt range (g)		Fish (n)		Survival (%)		Feed conversion ratio		Total food (%/body wt)	
	А	В	Α	В	Α	В	Α	В	Α	В	Α	В
Aug 06	246.72±21.62	250.11±21.98	195-285	200-300	250	250	100	100	2.46	2.23	8.20	7.43
Sep 06	442.61±26.00	450.27±24.79	400-490	400-500	250	250	100	100	1.16	1.10	3.86	3.66
Oct 06	592.16±25.66	600.00±27.66	545-648	550-650	250	250	100	100	1.85	1.81	6.16	6.03
Nov 06	742.22±27.61	750.00±28.78	695-800	700-800	250	250	100	100	2.35	2.28	7.83	7.60
Dec 06	984.72±34.01	1000.50±47.16	840-1146	850-1150	200	200	80 ¹	80 ¹	1.94	1.90	6.46	6.30
Jan 07	948.88±33.27	960.00±35.86	810-1070	860-1060	200	200	100	100	2.03	1.99	6.76	6.63
Feb 07	974.72±34.89	1000.05±64.94	820-1150	850-1190	200	200	100	100	2.21	2.15	7.36	7.10
Mar 07	1008.00±39.75	1025.00±35.35	900-1095	925-1100	200	200	100	100	2.08	2.02	6.93	6.73
Apr 07	1027.22±52.57	1050.00±40.00	925-1100	950-1100	200	200	100	100	1.96	1.93	6.53	6.43
May 07	1130.55±54.10	1150.00±53.74	1040-1250	1050-1250	200	200	100	100	1.97	1.95	6.56	6.50
Jun 07	1223.88±56.58	1250.00±55.27	1127-1305	1150-1300	188	192	93 ²	95 ²	1.91	1.90	6.36	6.33
Jul 07	1453.38±61.42	1478.16±57.84	1350-1582	1378-1578	188	192	100	100	1.89	1.85	6.30	6.16
Aug 07	1657.27±57.40	1676.00±54.76	1568-1774	1579-1779	188	192	100	100	1.87	1.86	6.23	6.20
Sep 07	1860.55±62.48	1877.00±56.20	1795-1965	1800-1954	188	192	100	100	1.91	1.90	6.36	6.33
Oct 07	2067.77±54.92	2095.00±53.85	1169-2198	1195-2195	188	192	100	100	1.92	1.89	6.40	6.30
Nov 07	2287.66±63.47	2313.05±56.23	2205-2421	2200-2426	188	192	100	100	1.95	1.90	6.50	6.33
Dec 07	2297.28±35.02 ^a	2508.38±41.31 ^b	2235-2520	2450-2567	188	192	100	100	2.04	1.95	6.80	6.50
Jan 08	2318.55±29.31	2483.00±25.91	2285-2525	2395-2530	188	192	100	100	2.16	1.99	7.20	6.63
Feb 08	2339.69±25.40	2594.22±21.80	2300-2600	2462-2610	188	192	100	100	2.25	2.06	7.50	6.86
<u>Mar 0</u> 8	2441.14±38.13 ^a	2785.00±64.91 ^b	2380-2693	2540-2820	188	192	100	100	1.91	2.03	6.36	6.86
Mean values in the same row with different superscripts significantly differ at $n < 0.05$												

Mean values in the same row with different superscripts significantly differ at p<0.05. ¹ mortality due to *Zeuxapta seriolae* infection

² mortality due to handling during monthly fish measurement



Month

Month

Fig. 3. (a) Daily live weight gain and (b) specific growth rate of Mediterranean amberjack (*Seriola dumerili*) fed commercial pellets for 20 months (Group A) or commercial pellets for 16 months and frozen sardines for 4 months (Group B).



Fig 4. Feed conversion ratios of Mediterranean amberjack fed commercial pellets (Group A) or pellets and sardines (Group B).

The FCR was low in comparison to other studies on amberjacks in the Mediterranean (Table 2). At the end of the feeding trial, fillets of the cultured amberjacks had significantly higher protein and lipid contents than those of wild by amberjacks captured local fishermen in the same area (Table 3). On the contrary, moisture and ash contents of the wild amberjacks were higher than those of their farmed counterparts (p<0.05).

Site	Cage system	Initial wt (g)	Final wt (g)	Cultivation period (days)	Diet	FCR	Reference
Castellammare, Sicily	Floating	130	670	85	Fish scraps	-	Giovanardi et al. 1984
Aeolian Islands, Sicily	Floating	73	858	120	Fish scraps	4.40	Porrello et al. 1993
Castellammare, Sicily	Submerged	48	576	120	Pellets	3.51	Mazzola et al. 2000
Castellammare, Sicily	Submerged	48	1129	120	Fish scraps	1.22	Mazzola et al. 2000
İskenderun Hatay, Turkey	Off-shore	102	2441	600	Pellets	2.01	This paper
İskenderun Hatay, Turkey	Off-shore	102	2785	600	Pellets & frozen sardines	1.93	This paper

Table 2. Comparison of results with other amberjack cage experiments in the Mediterranean.

Table 3. Proximate compositions (%) of farmed and wild amberjack, other fish species, and feeds.

	Moisture	Crude protein	Total lipid	Crude ash	Reference
Farmed S. dumerili	73.53±0.20	21.86±0.41	2.89±0.46	1.10 ± 0.06	This study
Wild captured S. dumerili	76.51±0.11	19.61±0.47	2.03±0.05	1.33 ± 0.07	This study
Sparus aurata	78.20±1.79	18.08±0.71	9.80±1.36	1.37 ± 0.08	Grigorakiş et al. 2003
Dicentrarchus labrax	70.30	21.80	7.69	1.32	Kyrana & Lougovois 2002
Commercial diet	9.25	45.00	22.50	9.00	-
Frozen sardine	75.83	20.05	0.99	3.04	Skaramuca et al. 2001

Discussion

The mortality rates were 24.8% in Group A and 23.2% in Group B. The high mortality resulted primarily from a *Zeuxapta seriolae* infection during December 2006, which accounted for 20% mortality. During the infection, the cages were subjected to formalin baths - a 300 ppm solution for 1 h during every week of infection. This treatment is typically used in aquaculture to exterminate monogeneans. The same procedure was followed before winter in 2007. In addition, the commercial diet was fortified by adding vitamin C. No further mortality occurred due to *Z. seriolae* infection after December 2006, indicating that the formalin bath treatments and vitamin C supplementation were effective together. The second incidence of mortality occurred during the monthly fish measurements in June 2007.

The wild captured Mediterranean amberjack juveniles adapted well to the cage conditions, accepting the provided food from stocking in the offshore culture system onwards. The growth rate was high throughout the 20 months of rearing except during winter. During adaptation to the commercial diet (July-September 2006), the fish in both groups accepted the food readily and attained higher specific growth rates than in other months. After adaptation, from winter 2007-8, group B reached a significantly larger size than group A, possibly because the amberjacks were observed to feed more readily on the frozen fish than on the commercial pellets.

Wide monthly changes in physico-chemical variables have a significant effect on amberjack growth in floating cages and tanks (Porrello et al., 1993). In our study, changes in growth performance and feed conversion ratios were closely related to water temperature. The growth performance can be divided into two distinct periods: April-November (high) and December-March (low). Different results were obtained in tanks by Garcia et al. (1993) and Skaramuca et al. (2001). The growth and maximum size of Group B were comparable to those obtained in floating cages (Giovanardi et al. 1984; Porrello et al., 1993) and tanks (Cavaliere et al., 1989; Greco et al., 1991). As expected, feed conversion ratios in both groups were higher in the winter than in the warmer months due to bad weather conditions and lower water temperatures. In general, FCR values were better than in earlier attempts conducted in the Mediterranean, except for those of Mazzola et al. (2000). Earlier reports support our findings that amberjack growth performance and daily rate of feeding decrease in autumn and winter (Mazzola et al., 2000; Skaramuca et al., 2001). During the high growth period, feed conversion rates were lower as long as the temperature was above 19°C. In the low growth period, when temperatures were relatively low (<19°C), the FCR rose. The difference between these two periods should be taken into account in amberjack aquaculture in the Mediterranean as these environmental conditions represent the real situation under which amberjack aquaculture will have to function.

The amberjacks in both groups had similar protein, lipid, ash, and moisture contents at the end of the experiment. Likewise, tank-reared Mediterranean amberjack fed pellets or frozen sardines had similar lipid, ash, moisture, and protein contents (Skaramuca et al. 2001). However, our amberjacks had a higher lipid content than the tank-reared amberjacks and significantly higher protein and lipid contents than wild amberjacks, indicating that Mediterranean amberjacks fed a commercial diet attain higher protein and lipid contents than wild and tank-reared specimens. Farmed Mediterranean amberjack are, thus, high-quality fish with a high percent of protein comparable to that of other farmed species and wild counterparts.

In conclusion, Mediterranean amberjack can be successfully reared in off-shore cage systems and has great aquaculture potential in the northeastern Mediterranean. In the winter, the juveniles consumed the frozen sardines more readily than the commercial feed, thus it is advised to use frozen sardines and/or semi-moist pelleted diets during the winter grow-out period for better feed intake and growth performance. Further research is certainly needed for sustainable culture of Mediterranean amberjack especially in the following areas: feeding strategy, reproductive control, sustainable larval rearing, and nutritional requirements.

Acknowledgements

This work was funded by the Scientific and Technological Research Council of Turkey, Project No. 105 O 510.

References

Andaloro F., Potoschi A. and S. Porrello, 1992. Contribution to knowledge of growth of greater amberjack, *Seriola dumerili* (Cuv. 1817) in the Sicilian Channel (Mediterranean Sea). *Rap. Int. Mer Med*, 33:282.

AOAC, 1990. *Official Methods of Analysis*, 15th ed. Assoc. of Official Analytical Chemists, Arlington, VA. 1298 pp.

Bligh E.G., and W.J. Dyer, 1959. A rapid method of total lipid extraction and purification. *Can. J. Biochem. Physiol.*, 37:911-917.

Cavaliere A., Crisafi E., Faranda F., Greco S., Lo Paro G., Manganaro A. and A. Mazzola, 1989. Collection of fingerling and rearing of *Seriola dumerili* in tanks. pp. 119-123. In: N. De Pauw, E. Jaspers, H. Ackefors, N. Wilkins (eds.). *Aquaculture - A Biotechnology in Progress*. N. Eur. Aquacult. Soc., Bredene, Belgium.

Fischer W., 1978. *FAO Species Identification Sheets for Fishery Purposes, Western Central Atlantic (Fishing Area 31)*. Marine Resources Services, Fishery Resources and Environment Division, Fish. Dept., FAO, Rome.

Garcia B., Moreno J. and M.J. Rosique, 1993. Rate of feeding, growth and gross efficiency in juveniles of yellowtail, *Seriola dumerili*. pp. 255-260. In: *Production, Environment and Quality.* Eur. Aquacult. Soc. Spec. Publ. No. 18.

Giovanardi O., Mattioli G., Piccinetti C. and G. Smabucci, 1984. Prime esperienze sull'allevamento della Ricciola (*Seriola dumerili*, Risso 1810) in Italia. *Riv. It. Piscic Ittiop.* A-XIX, 4:123-130.

Greco S., Arena G., Caridi D. and V. Micale, 1991. An improved method of capture and transport for juveniles of *Seriola dumerili*. In: N. De Pauw, J. Joyce (eds.). *Aquaculture and Environment. Dublin Aquacult.*, 14:130-131.

Grigorakiş K., Taylor K.D.A. and M.N. Alexis, 2003. Organoleptic and volatile aroma compounds comparison of wild and cultured gilthead sea bream (*Sparus aurata*): sensory differences and possible chemical basis. *Aquaculture*, 225:105-119.

Jover M., Garcia-Gomez A., Tomas A., De la Gandara F. and L. Perez, 1999. Growth of the Mediterranean yellowtail (*Seriola dumerilii*) fed extruded diets containing different levels of protein and lipid. *Aquaculture,* 179:25-33. **Kyrana V.R. and V.P. Lougovois,** 2002. Sensory, chemical and microbiological assessment of farm-raised European sea bass (*Dicentrarchus labrax*) stored in melting ice. *Int. J. Food Sci. Technol.*, 37:319-328.

Laroche W.A., Smith-Vaniz W.F. and S.L. Richardson, 1984. Carangidae: development. pp. 510-522. In: H.G. Moser (ed.). *Ontogeny and Systematics of Fishes*. Spec. Publ. No. 1. Am. Soc. Ichthyol. Herpetol., Allen Press, Lawrence, KA.

Lazzari A., Fusari A., Boglione A., Marino G. and M. Di Francesco, 2000. Recent advances in reproductional and rearing aspects of *Seriola dumerilii*. pp. 241-247. In: B. Basurco (ed.). *Cahiers Options Mediterraneennes, Mediterranean Marine Aquaculture Finfish Species Diversification*, vol. 47. CIHEAM, Zaragoza, Spain.

Marino G., Mandich A., Massari A., Andaloro F. and S. Porrello, 1995. Aspects of reproductive biology of the Mediterranean amberjack (*Seriola dumerilii* Risso, 1810): gonadal development. pp. 115-124. In: B. Basurco (ed.). *Cahiers Options Mediterraneennes, Marine Aquaculture Finfish Species Diversification*, vol. 16. CIHEAM, Zaragoza, Spain.

Mazzola A., Favaloro E. and G. Sara, 2000. Cultivation of the Mediterranean amberjack, *Seriola dumerili* (Risso, 1810), in submerged cages in the western Mediterranean Sea. *Aquaculture*, 181:257-268.

Nakada M., 2000. Yellowtail and related species culture. pp. 1007-1036. In: R.R. Stickney (ed.). *Encyclopedia of Aquaculture*. Wiley, London.

Pastor E., Grau A., Riera F., Pou S., Massuti E. and A.M. Grau, 2000. Experiences in the culture of new species in the Estacion de Acuicultura of the Balearic Government (1980–1998). pp. 371-379. In: B. Basurco (ed.). *Cahiers Options Mediterraneennes, Marine Aquaculture Finfish Species Diversification*, vol. 16. CIHEAM, Zaragoza, Spain.

Porrello S., Andaloro F., Vivona P. and G. Marino, 1993. Rearing trial of *Seriola dumerili* in a floating cage. pp. 299-307. In: *Production, Environment and Quality.* Eur. Aquacult. Soc. Spec. Publ. No. 18.

Skaramuca B., Kozul V., Teskeredzic Z.J., Bolotin J. and V. Onofri, 2001. Growth rate of tank-reared Mediterranean amberjack, *Seriola dumerili* (Risso 1810) fed on three different diets. *J. Appl. Ichthyol.*, 17(3):130-133.

Tachihara K., Ebisu R. and Y. Tukashima, 1993. Spawning, eggs, larvae and juveniles of the purplish amberjack (*Seriola dumerili*). *Nippon Suisan Gakkaishi*, 59(9):1479-1788.

Takakuwa F., Fukada H., Hosokawa H. and T. Masumoto, 2006. Optimum digestible protein and energy levels and ratio for greater amberjack *Seriola dumerili* (Risso) fingerling. *Aquacult. Res.*, 37:1532-1539.

Thompson B.A., Beasly M. and C.A. Wilson, 1999. Age distribution and growth of greater amberjack *Seriola dumerili*, from the north-central Gulf of Mexico. *Fish. Bull.*, 97:362-371.

Zar H., 1984. *Biostatistical Analyses*. Prentice-Hall, NJ.