

The Open Access Israeli Journal of Aquaculture – Bamidgeh

As from **January 2010** The Israeli Journal of Aquaculture - Bamidgeh (IJA) will be published exclusively as **an on-line Open Access (OA)** quarterly accessible by all AquacultureHub (<http://www.aquaculturehub.org>) members and registered individuals and institutions. Please visit our website (<http://siamb.org.il>) for free registration form, further information and instructions.

This transformation from a subscription printed version to an on-line OA journal, aims at supporting the concept that scientific peer-reviewed publications should be made available to all, including those with limited resources. The OA IJA does not enforce author or subscription fees and will endeavor to obtain alternative sources of income to support this policy for as long as possible.

Editor-in-Chief

Dan Mires

Editorial Board

Rina Chakrabarti	Aqua Research Lab, Dept. of Zoology, University of Delhi, India
Angelo Colorni	National Center for Mariculture, IOLR Eilat, Israel
Daniel Golani	The Hebrew University of Jerusalem Jerusalem, Israel
Hillel Gordin	Kibbutz Yotveta, Arava, Israel
Sheenan Harpaz	Agricultural Research Organization Beit Dagan,
Gideon Hulata	Agricultural Research Organization Beit Dagan,
George Wm. Kissil	National Center for Mariculture, IOLR, Eilat, Israel
Ingrid Lupatsch	Swansea University, Singleton Park, Swansea, UK
Spencer Malecha	Dept. of Human Nutrition, Food & Animal Sciences, CTAHR, University of Hawaii
Constantinos Mylonas	Hellenic Center for Marine Research, Crete, Greece
Amos Tandler	National Center for Mariculture, IOLR Eilat, Israel
Emilio Tibaldi	Udine University Udine, Italy
Jaap van Rijn	Faculty of Agriculture, The Hebrew University of Jerusalem, Israel
Zvi Yaron	Dept. of Zoology, Tel Aviv University, Tel Aviv, Israel

Published under auspices of
**The Society of Israeli Aquaculture and
Marine Biotechnology (SIAMB),
University of Hawai'i at Mānoa Library**

&

**University of Hawai'i at Mānoa
Aquaculture Program**
in association with
AquacultureHub

<http://www.aquaculturehub.org>



UNIVERSITY
of HAWAII
MĀNOA
LIBRARY



AquacultureHub
educate • learn • share • engage

ISSN 0792 - 156X

© Israeli Journal of Aquaculture - BAMIGDEH.

PUBLISHER:

Israeli Journal of Aquaculture - BAMIGDEH -
Kibbutz Ein Hamifratz, Mobile Post 25210,
ISRAEL

Phone: + 972 52 3965809

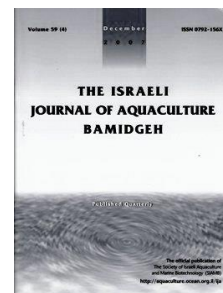
<http://siamb.org.il>

Copy Editor **Ellen Rosenberg**



The *IJA* appears exclusively as a peer-reviewed on-line open-access journal at <http://www.siamb.org.il>. To read papers free of charge, please register online at [registration form](#).

Sale of *IJA* papers is strictly forbidden.



DNA Barcoding Analysis of Commercial Freshwater Fish Species Cultured in China

Qian Wang¹, Wei Dai¹, Xiaomei Wang¹, Jianxin Yang², Wenli Zhou^{1*}

¹ *Tianjin Key Laboratory of Aqua-Ecology and Aquaculture, Department of In line Fisheries Science, Tianjin Agricultural University, Tianjin 300384, P.R. China*

² *Tianjin Huanxin Seed Multiplication Farm of freshfish, Tianjin 300384, P.R. China*

(Received 29.1.2015, Accepted 17.3.2015)

Key words: DNA barcoding, freshwater fish, COI, hybrid compatibility, genetic distance,

Abstract

In this study, thirteen commercial freshwater fish species cultured in China, and one hybrid F1 (*Megalobrama amblycephala* ♀ × *Erythroculter ilishaeformis* ♂), were barcoded for a 860 bp region of the mitochondrial cytochrome oxidase subunit I (COI) gene. The results showed that a neighbor-joining (NJ) tree was constructed based on COI sequence clustered species in accordance with their taxonomic classification. Hybrid F1 could not be differentiated from *M.amblycephala* in the neighbor-joining (NJ) tree. There was a significant negative correlation between interspecific hybrid compatibility and genetic distance ($R^2 = 0.96$). Hybrid compatibility can be predicted by the interspecific genetic distance.

Introduction

Interspecific hybridization programs have been applied in fish farms with the purpose of producing animals that perform better than the parental species (hybrid vigor) (Bartley et al., 2001). However, the hybrid incompatibility caused by reproductive isolation always impedes interspecific hybridization. There is a correlation between parental divergence and reproductive isolation (Edmands, 2002). If interspecific hybrid compatibility could be predicted based on the genetic divergence of parents, this would be of great use to fish breeders. Genetic distance, representing interspecific genetic divergence, is often used and is a popular tool for species classification and evolutionary studies (Shirak et al., 2009; Wang et al., 2001) using molecular markers, including DNA barcoding.

DNA barcoding uses sequence diversity within a short and standardized gene region as a molecular diagnostic tool for species-level identification (Toffoli et al., 2008). DNA barcode sequences are very short relative to the entire genome and can be obtained reasonably quickly, and cheaply (Ardura et al., 2010). Hebert et al. (2003) proposed that a single gene sequence would be sufficient to differentiate all, or at least the vast majority of, animal species. The mitochondrial cytochrome oxidase subunit I (COI) gene has been employed as a global bioidentification sequence to catalogue the world's animal taxa, including fish (Ward et al., 2008). Combined with the potential for automated, rapid sample processing (Garland and Zimmer 2002), DNA barcoding could soon provide a powerful foundation for accurate and unambiguous fish species identification from eggs to adults, as well as analysis of phylogenetic relationship (Ward et al., 2009; Ardura et al., 2010; Zhang and Hanner, 2011). Moreover, it provides a tool for authentication of commercial fish products like fish slices and fillets that could not be morphologically identified, (Galal-Khallaf et al., 2014).

In this study, we examined sequence divergence at COI in 13 freshwater fish species and one hybrid. All of these are important commercial species cultured in China. The investigation may provide a further test of COI barcode for freshwater fish identification.

Materials and Methods

Fish specimens and DNA extraction. One hybrid F1 (*M.amblycephala* ♀ × *E. ilishaeformis* ♂) and 11 freshwater fish species belonging to 10 genera, 3 families, and 2 orders were provided by Tianjin Huanxin Seed Multiplication Farm of fresh fish, Tianjin, China (Table 1).

Table1. List of analyzed specimens with associated sample.

Species	Genera	Families	Orders
<i>Aristichthys nobilis</i>	<i>Aristichthys</i>	Cyprinidae	Cypriniformes
<i>Carassius auratus pengze</i> (PZ)	<i>Carassius</i>	Cyprinidae	Cypriniformes
<i>Carassius auratus red var</i> (R)			
<i>Carassius curatus cuvieri</i> (W)	<i>Carassius</i>	Cyprinidae	Cypriniformes
<i>Ctenopharyngodon idellus</i>	<i>Ctenopharyngodon</i>	Cyprinidae	Cypriniformes
<i>Cyprinus carpio</i> (G)	<i>Cyprinus</i>	Cyprinidae	Cypriniformes
<i>Cyprinus carpio</i> (BL)			
<i>Cyprinus carpio haematopterus</i> (WK)			
<i>Cyprinus carpio specularis</i> (KL)			
<i>Erythroculter ilishaeformis</i>	<i>Erythroculter</i>	Cyprinidae	Cypriniformes
<i>Hypophthalmichthys molitrix</i>	<i>Hypophthalmichthys</i>	Cyprinidae	Cypriniformes
<i>Ictiobus cyprinellus</i>	<i>Ictiobus</i>	Catostomidae	Cypriniformes
<i>Megalobrama amblycephala</i>	<i>Megalobrama</i>	Cyprinidae	Cypriniformes
<i>Myxocyprinus asiaticus</i>	<i>Myxocyprinus</i>	Catostomidae	Cypriniformes
<i>Oreochromis niloticus</i>	<i>Oreochromis</i>	Cichlidae	Perciformes

Abbreviations of subspecies name used in China are in parentheses.

WK: Russian carp; G: Germany mirror carp; BE: Birnir carp; KL: mirror carp; W: Japanese crucian carp; R: red crucian carp; PZ: Pengze carp.

Total sample size was 88 specimens. Tissue samples from each of the specimens were collected aseptically and preserved in 95% ethanol. A small piece of alcohol-preserved tissue was dissolved in 200 µL of STE buffer (1mM Tris HCl, 1mM EDTA and 10 mM NaCl) in a microcentrifuge tube. Total DNA was extracted using Phenol-Chloroform-

Isoamylalcohol method. The DNA samples were then dissolved in nuclease-free water and stored at -20°C until further use.

PCR amplification and sequencing. The COI gene (860 bp) was amplified using the set of published primers (Peng et al., 2009) as follows: L5956-COI (5'-CACAAAGACATTGGCACCCT-3') and H6855-COI (5'-AGTCAGCTGAAKACTTTTAC-3'). The amplification reactions were performed in a total volume of 50 µl comprising 1X PCR buffer, 2.5 mM MgCl₂, 0.5µM of each primer, 0.25 mM of each dNTPs(TAKARA™), 1-2 U of Taq polymerase (TAKARA™) and 100 ng of DNA template. The thermocycler conditions for the amplification were: an initial denaturation at 94°C for 4 min, then 35 cycles of denaturation at 94°C for 45 s, annealing at 48°C for 30 s and extension at 72°C for 1 min 30 s, followed by a final extension at 72°C for 5 min. The PCR-amplified products were analyzed in 1.8% agarose gels. Those products with a single uniform band were purified, and then sequenced in an automated DNA sequencer (ABI 3730, Applied Biosystems Inc., CA, USA) at Sangon Biotech(Shanghai) Co., Ltd.

Analyzing correlation between hybrid compatibility and genetic divergence. According to the definition of Xu et al. (2010), hybrid compatibility was scored as follows: 1, both negative and positive cross F1 viable; 0.5, either negative or positive cross F1 viable; 0, both negative and positive cross F1 inviable. Interspecific hybridizations among some freshwater fish species investigated in this study have been performed by Tianjin Huanxin Seed Multiplication Farm of freshfish (Tianjin, China) (Jin, 2009). Interspecific hybrid compatibility among 8 freshwater fish species reported by Jin (2009) and genetic distance estimated using COI barcodes are summarized in Table 2. The linear correlation between hybrid compatibility and genetic divergence (genetic distance) was analyzed using Excel 2007.

Table 2. K2P genetic distance (%) within various taxonomic levels.

Comparisons within	Numbers of comparisons	Genetic distance (%)				
		Taxa	Minimum	mean	maximum	SE
Species	84	13	0	0.34	5.5	0.009
Genera	84	10	0.1	5.00	12.6	0.018
Families	84	3	0.5	14.5	19.9	0.015
Orders	84	2	17.1	32.2	63.6	0.028

Data analyses. Two COI sequences of *Cyprinus pellegrini* and *Cyprinus multitaeniata* were referenced from the Genbank (accession numbers: JX042166 and HM536896). All sequences we sequenced and referenced were aligned using Clustal X 1.8 software. Sequence divergences were calculated using the Kimura two parameter (K2P) genetic distance model (Kimura 1980). Neighbour-joining (NJ) tree of K2P genetic distance was created to provide a graphic representation of the patterning of divergence between species (Saitou and Nei 1987). Bootstrapping was performed in MEGA 4.0 (Tamura et al. 2007) with 1000 replications.

Results

The average K2P genetic distance of individuals within species was 0.34% compared with 5.00 % for species within genera (table 2). There was 15-fold more variation among congeneric species than among conspecific individuals. Mean divergence among species within families and classes increased to 14.5% and 32.2%, respectively (table 1). Overall nucleotide frequencies of COI sequence were C (26.4%), T (29.1%), A (26.2%), G (18.4%), respectively.

As showed in the NJ tree (Fig. 1), *O. niloticus* which showed the greatest genetic divergence was placed at the base of the tree. The different genera were derived and formed branches respectively. All COI sequences formed species units were clustered in monophyletic groups at the genus level. Hybrid F1 (*M.amblycephala* ♀ × *E.ilishaeformis* ♂) could not be differentiated from *M.amblycephala* in the NJ tree. Genetic distance between hybrid F1 and *M.amblycephala* ♀ (0.2%) was much closer than that between hybrid F1 and *E.ilishaeformis* ♂ (6.1%). This indicated that hybrid F1 had no hereditary similarity.

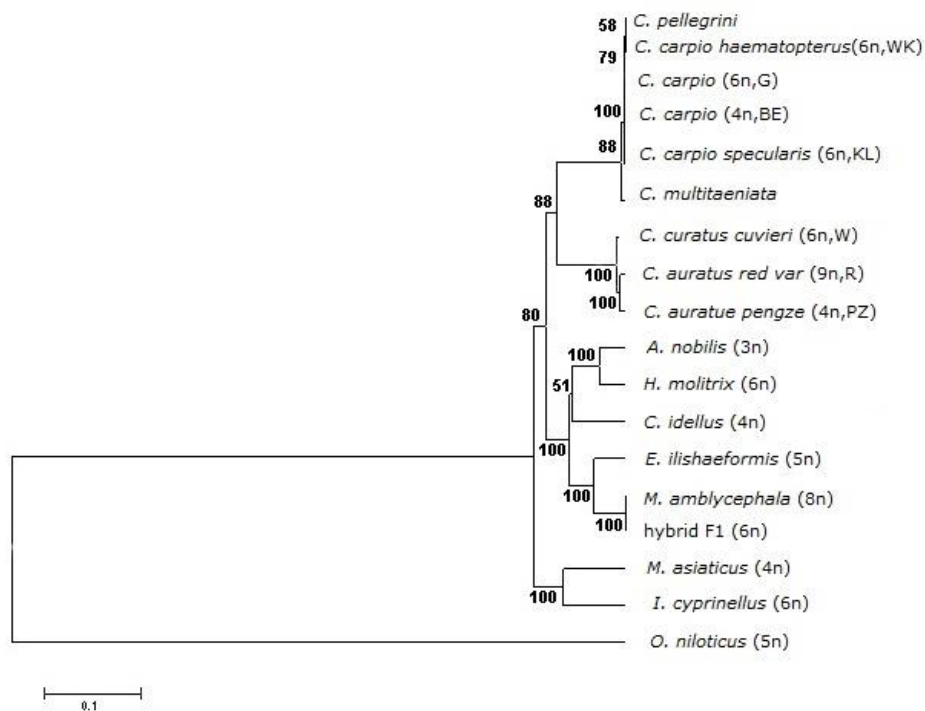


Fig. 1. Neighbor-joining tree of 90 COI sequences from 13 freshwater fish species and one hybrid F1 (*M.amblycephala* ♀ × *E. ilishaeformis* ♂), using K2P distances. In parentheses are the numbers of specimens sequenced.

As shown in Fig. 2, there was a significant negative correlation between interspecific hybrid compatibility and genetic distance ($R^2 = 0.96$). Both negative and positive cross F1 were viable when genetic distance was equal to 6.1 (table 3).

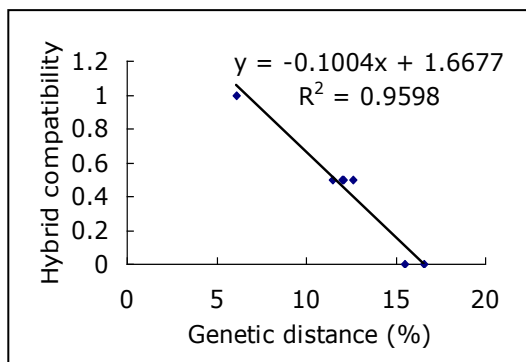


Fig. 2. The correlation between hybrid compatibility and genetic distance of freshwater fish species.

Table 3. Hybrid compatibility and K2P genetic distance among 8 freshwater fish species.

A	B	A×B	B×A	Hybrid compatibility	Genetic distance (%)
<i>M. amblyceph</i>	<i>E. ilishaeformis</i>	1	1	1	6.1
<i>A. nobilis</i>	<i>H. molitrix</i>	1	0	0.5	11.5
<i>C. carpio</i>	<i>C. auratus red var</i>	1	0	0.5	12.0
<i>haematopterus</i>					
<i>C. carpio specularis</i>	<i>C. auratus red var</i>	1	0	0.5	12.1
<i>C. curatus cuvieri</i>	<i>C. carpio</i>	1	0	0.5	12.6
<i>C. auratus red var</i>	<i>M. amblyceph</i>	0	0	0	15.5
<i>I. cyprinellus</i>	<i>C. auratus red var</i>	0	0	0	16.6

Discussion

We found the guanine-cytosine (GC) content of the COI region in 13 freshwater fish species of Osteichthyes was 44.8 %. The GC contents in 143 and 9 Osteichthyes species were 47.1% and 43.2%, respectively (Ward et al. 2005; Saccone et al. 1999). Higher GC content (above 40%) in mitochondrial genome might be a genetic feature of teleost.

A threshold to delimit species with DNA barcode data has been proposed (Hebert et al 2004a,b). At least 10 times the average intraspecific distance should be flagged as provisional new species. We found the average genetic distance within conspecific specimens (0.34 %) was 15 times lower than that found among congeneric species (5.00 %). These values were consistent with those found in freshwater fish from Canada (0.27% (conspecific) and 8.37% (congeneric)) (Hubert et al., 2008), Mexico and Guatemala (0.45% (conspecific) and 5.1% (congeneric)) (Valdez-Moreno et al., 2009),

Cuba (0.6% (conspecific) and 9.1% (congeneric)) (Lara et al., 2010). All these data suggest that DNA barcoding can be used as a useful tool for the identification of freshwater fish species.

Hybrid F1 (*M.amblycephala* ♀ × *E. ilishaeformis* ♂) could not be differentiated from *M.amblycephala* in NJ tree constructed with the COI barcode. Hybridization among species would create taxonomic uncertainty when mitochondrial DNA is maternally inherited because any hybrid or subsequent generation would have the maternal DNA only (Ward et al., 2005). Where species boundaries were blurred by hybridization, supplemental analyses of one or more nuclear genes will be required.

The positive relationship between reproductive isolation and genetic distance has been confirmed in angiosperm (Moyle et al., 2004), butterflies (Presgraves, 2002), birds (Lijtmaer et al. 2003), and fish (Russell 2003; Bolnick and Near, 2005). Consistent with above investigation results, hybrid compatibility among freshwater fishes decreased with the increasing genetic distances. To improve the success rate of interspecific hybridization, hybrid compatibility should be predicted first by assessing the genetic distances of the parents.

Acknowledgements

The financial support provided by the Natural Science Foundation of Tianjin (Grant No.11JCYBJC08200), Program for Tianjin Innovative Research Team in University (Grant No. TD12-5018) and Key Projects of Applied Basic and Frontier technology Research of Tianjin (Grant No. 13JCZDJC29300) are gratefully acknowledged.

References

- Ardura A., Linde A.R., Moreira J.C. and E. Garcia-Vazquez**, 2010. DNA barcoding for conservation and management of Amazonian commercial fish. *Biol. Conserv.*, 143 (6): 1438-1443.
- Bartley D.M., Rana K., and A.J Immink**, 2001. The use of inter-specific hybrids in aquaculture and fisheries. *Rev. Fish Biol. Fish.*, 10: 325-337.
- Bolnick D.I. and T.J. Near**, 2005. Tempo of hybrid inviability in centrarchid fishes (Teleostei: *Centrarchidae*). *Evolution*, 59(8):1754-1767.
- Edmands S.**, 2002. Does parental divergence predict reproductive compatibility? *Trend. Ecol. Evol.*, 17:520-527.
- Galal-Khallaf A., Ardura A., Mohammed-Geba K., Borrell Y.J. and E. Garcia-Vazquez**, 2014. DNA barcoding reveals a high level of mislabeling in Egyptian fish fillets. *Food Control*, 46:441-445.
- Garland E.D. and C.A. Zimmer**, 2002. Techniques for the identification of bivalve larvae. *Mar. Ecol. Prog. Ser.*, 225: 299-310.
- Hebert P.D.N., Cywinska A., Ball S.L. and J.R. deWaard**, 2003. Biological identifications through DNA barcodes. *Proc. R. Soc. B.*, 270: 313-321.
- Hebert P.D.N., Penton E.H., Burns J.M., Janzen D.H. and W. Hallwachs**, 2004a. Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly *Astraptes fulgerator*. *Proc. Natl. Acad. Sci. USA*, 10(41):14812-14817.
- Hebert P.D.N., Stoeckle M.Y., Zemplak T.S. and C.M. Francis**, 2004b. Identification of birds through DNA barcodes. *PLoS. Biol.*, 2(10): 1657-1663.
- Hubert N., Hanner R., Holm E., Mandrak N.E., Taylor E., BurrIDGE M., Watkinson D., Dumont P., Curry A., Bentzen P., Zhang J., April J. and L. Bernatchez**, 2008. Identifying Canadian freshwater fishes through DNA barcodes. *PLoS. ONE.*, 3(6):e2490.
- Jin W.K.**, 2009. *Experimental report of distant hybridization in freshwater fish*. Chinese Agricultural Science and Technology Press, Beijing.
- Kimura M.**, 1980. A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *J. Mol. Evol.*, 15:111-120.
- Lara A., Ponce de Leon J.L., Rodriguez R., Casane D., Cote G., Bernatchez L. and E. Garcia-Machado**, 2010. DNA barcoding of Cuban freshwater fishes: Evidence for cryptic species and taxonomic conflicts. *Mol. Ecol. Resour.*, 10(3):421-430.

- Lijtmaer D.A., Mahler B. and P.L. Tubaro**, 2003. Hybridization and postzygotic isolation patterns in pigeons and doves. *Evolution*, 57(6):1411–1418.
- Presgraves D.C.**, 2002. Patterns of postzygotic isolation in Lepidoptera. *Evolution*, 56(6):1168–1183.
- Moyle L.C., Olson M.S. and P. Tiffin**, 2004. Patterns of reproductive isolation in three angiosperm genera. *Evolution*, 58(6): 1195–1208.
- Peng J.L., Wang X.Z., Wang D. and S.P. He**, 2009. Application of DNA barcoding based on the mitochondrial COI gene sequences in classification of Culter (Pisces: Cyprinidae). *Acta Hydrobiologica Sinica*(in Chinese), 33(2):271–276.
- Russell S.T.**, 2003. Evolution of intrinsic post-zygotic reproductive isolation in fish. *Ann. Zool. Fenn.*, 40:321–329.
- Saccone C., De Giorgi C., Gissi C., Pesole G. and A. Reyes**, 1999. Evolutionary genomics in Metazoa: the mitochondrial DNA as a model system. *Gene*, 238(1):195–209.
- Saitou N. and M. Nei**, 1987. The neighbour-joining method: a new method for reconstructing evolutionary trees. *Mol. Biol. Evol.*, 4: 406–425.
- Shirak A., Cohen-Zinder M., Barroso R.M., Eyal Seroussi E., Ron M. and Hulata G.**, 2009. DNA barcoding of Israeli indigenous and introduced cichlids. *The Israeli Journal of Aquaculture – Bamidgeh* 61(2), 83–88.
- Tamura K., Dudley J., Nei M. and S. Kumar**, 2007. MEGA4: molecular evolutionary genetics analysis (MEGA) software version 4.0. *Mol. Biol. Evol.*, 24: 1596–1599.
- Toffoli D., Hrbek T., de Araujo M.L.G., de Almeida M.P., Charvet-Almeida P. and I.P.Farias**, 2008. A test of the utility of DNA barcoding in the radiation of the freshwater stingray genus *Potamotrygon* (Potamotrygonidae, Myliobatiformes). *Genet. Mol. Biol.*, 31: 324–336.
- Valdez-Moreno M., Ivanova N.V., Elías-Gutiérrez M., Contreras-Balderas S. and P. D. N. Hebert**, 2009. Probing diversity in freshwater fishes from Mexico and Guatemala with DNA barcodes. *J. Fish. Biol.*, 74(2):377–402.
- Wang R., Zheng L.B., Toure Y.T., Dandekar T. and F.C. Kafatos**, 2001. When genetic distance matters: Measuring genetic differentiation at microsatellite loci in wholegenome scans of recent and incipient mosquito species. *Proc. Natl. Acad. Sci. USA*, 98(19): 10769–10774.
- Ward R.D., Costa F.O., Holmes B.H. and D. Steinke**, 2008. DNA barcoding of shared fish species from the North Atlantic and Australasia: minimal divergence for most taxa, but *Zeus faber* and *Lepidopus caudatus* each probably constitute two species. *Aquat. Biol.*, 3: 71–78.
- Ward R.D., Hanner R. and P.D.N. Hebert**, 2009. The campaign to DNA barcode all fishes, FISH-BOL. *J. Fish Biol.*, 74: 329–356.
- Ward R.D., Zemlak T.S., Innes B.H., Last P.R. and P.D. Hebert**, 2005. DNA barcoding Australia's fish species. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.*, 360(1462):1847–1857.
- Xu D.D., You F., Lou B., Li J., Xu J.H., Wu Z.H. and P.J. Zhang**, 2010. Analysis of correlation between pairwise genetic distance of eight flatfishes and hybrid fitness. *Journal of fisheries of China*(in Chinese), 34(2):178–184.
- Zhang J.B. and R. Hanner**, 2011. DNA barcoding is a useful tool for the identification of marine fishes from Japan. *Biochem. Syst. Ecol.*, 39 (1): 31–42.